Total Synthesis of the Proposed Structure for Aromin and Its Structural Revision

Shunya Takahashi,^{*,†} Daisuke Satoh,[†] Misato Hayashi,[†] Kohta Takahashi,[‡] Kazunori Yamaguchi,[§] Takemichi Nakamura,[†] and Hiroyuki Koshino[†]

[†]RIKEN Center for Sustainable Resource Science, Wako, Saitama 351-0198, Japan

[‡]Division of Cell Recognition Study, Institute of Molecular Biomembrane and Glycobiology, Tohoku Medical and Pharmaceutical University, Sendai 981-8558, Japan

[§]Division of Molecular and Cellular Oncology, Miyagi Cancer Center Research Institute, Natori 981-1293, Japan

Supporting Information



ABSTRACT: This paper describes the first total synthesis of the proposed structure for aromin, an annonaceous acetogenin possessing an unusual bis-THF ring system, and its 4*S*,7*R*-isomer. The key steps involve an oxidative cyclization of a couple of terminal-diene alcohols and an intermolecular metathesis of an alkenyl tetrahydrofuran with an enone carrying a tetrahydrofuranyl lactone. The spectral data of both samples did not match those of aromin. Re-examination of the NMR data using the CAST/CNMR Structure Elucidator and chemical derivations suggested that the real structure of aromin should be revised to be a tetrahydropyran acetogenin, montanacin D. Cytotoxicities in human solid tumor cell lines for synthetic samples were also evaluated.

■ INTRODUCTION

The Annonaceous acetogenins from Annonaceae plants comprise a class of almost 420 natural products that exhibit a remarkably broad spectrum of biological properties such as anticancer, antiinfective, immunosuppressive, antifeedant, and pesticidal activities. Structurally, most of these compounds are characterized by a terminal γ -lactone unit at the end of a long aliphatic chain containing one to three tetrahydrofuran (THF) rings or tetrahydropyran (THP) ring or epoxide rings, or other functional groups.¹ Certain acetogenins possess an unusual structure forming a cyclic ether by a 4-hydroxy group adjacent to the γ -lactone ring.^{2–4} In 1996, McLaughlin et al. isolated a couple of new acetogenins, aromin and aromicin from the stem bulk of *Xylopia aromatica*.⁵ Their structures were elucidated by chemical and spectral means to be 1 and 2 possessing a 4,7trans THF ring along with a 16,19-trans THF ring as a common scaffold, respectively (Figure 1). The trans configuration of both THF rings in 1 was deduced by the relatively large δ



Figure 1. Proposed structures for aromin (1) and aromicin (2).

differences between the gem-protons in the THF rings and by no cross-peak at H-4/H-7 or at H-16/H-19 in the NOESY spectrum;^{6,7} the relative stereochemistry of the $C_{15}-C_{20}$ portion was also confirmed by comparison of their ¹³C NMR data with those of model compounds.⁸ The absolute configurations of C-15 and C-20 in 1 were determined by the Mosher method while the stereochemistries at C-4 and C-34 (C-36 for 2) were shown to be 4R, 34S by comparison of the CD curve of 1 with those of several acetogenins previously proven to have 4R, 34S configurations. Since 1 and 2 differ only by the length of the carbon chain, 2 was assumed to have the same absolute stereochemistry. Both natural products showed significant cytotoxicities among six human tumor cell lines; however, the activity was notably reduced compared to other nonadjacent bis-THF ring acetogenins. We presumed that the reason would be due to a conformational rigidity around the lactone ring, an essential domain for several biological activities. In connection with our synthetic studies on Annonaceous acetogenins,⁹ the unique structure stimulated our interest. Described herein is the first total synthesis of aromin that dictates revision of the formula of 1 to 32 (montanacin D).

Received: September 8, 2016 Published: November 4, 2016

RESULTS AND DISCUSSION

Our synthetic strategy directed toward 1 was based on a convergent process that involved intermolecular metathesis of olefin 3 and enone 4 as illustrated in Scheme 1, and we recently

Scheme 1. Synthetic Plan of 1



demonstrated the usefulness of this method.^{9a,b} The enone 4 might be prepared from γ -lactone 6^{10} and a THF derivative 7 while the left-half segment 3 would be synthesized from a THF alcohol 5. The presence of the *trans*-THF ring system in both parts made us envisage an oxidative cyclization¹¹⁻¹³ of terminal diene diols 8 and 9. Since no paper has appeared dealing with the oxidative cyclization of such diene alcohols,^{14–16} this strategy might be more challenging.

Synthesis of the left-half segment corresponding to the C₁₀- C_{32} domain began with a chain elongation reaction¹⁷ of 10^{18} (Scheme 2). After hydrolysis of the isopropylidene group in 11, the resulting diol 8 was treated with 10 mol % of tert-butyl hydroperoxide in the presence of $Co(nmp)_2$ (10 mol %) in 2propanol under an oxygen atmosphere¹³ to give a *trans*-THF alcohol 5^{19} in good yield. As expected, a tetrahydropyran²⁰ or an oxepan derivative and a bicyclic compound including such ring systems were not isolated. Hydroxyl protection of 5 with TBSOTf followed by selective deprotection gave a primary alcohol 12. After Dess-Martin oxidation, the resulting aldehyde reacted with dodecylmagnesium bromide to provide a desired threo alcohol 14 in 73% yield along with its epimer 13 (18%). Each relative stereochemical relationship between the chiral centers at C-10/C-11 was defined based on the NMR data,^{7,} and the stereoselectivity would be explained by an α -chelation controlled reaction pathway. Treatment of 14 with TBSOTf gave the left-half segment 3.

Having completed the synthesis of the left-half segment, we next turned our attention to preparation of the right-half segment. The fact that the absolute configuration of the THF ring in the segment was not confirmed prompted us to prepare both enantiomers of the part. Prior to the experiment, an oxidative cyclization of 1,3-*anti*-diol **9a** and its *syn* isomer **9b** was considered.²¹ Different from the cyclization of **8**, there are 4 types of possible pathways (Scheme 3). In the case of **9a**, we





predicted that the Co-olefin complex interacting with a C-3 alkoxy radical would cause a conformational lock of the vinyl moiety, resulting in inhibition of path a. On the other hand, path d leading to 18 from 9b may be unfavorable because there should be a serious repulsion in the transition state. Therefore, allyl alcohol such as 7a or 7b was expected to be a major product in both cases. An initial attempt to prepare the proposed structure 7a from 9a was conducted under Mukaiyama's conditions,¹² affording a single product as judged by TLC analysis. However, it was revealed to be an inseparable mixture of two compounds by ¹H NMR spectra. Each compound could be separated after acetylation and characterized to be 16 and 17 (Table 1).^{19,28} As expected, the major product was 17 through path b. The modified procedure reported by Pagenkopf et al.¹³ slightly improved the yield of 17 (57%) but 16 was also obtained in 26% yield. Upon deacetylation, 17 afforded the desired diol 7a. The cyclization of 1,3-syn-diol 9b also resulted in an inseparable mixture of cyclized products, and after acetylation, THF derivatives 19 and 20 were separated (Table 1). Compound 19 was transformed into 7b in a similar way. As the regioselectivity in the cyclization resulted in being not as high as expected, we decided to develop a more efficient route to the THF core.²⁹

The second approach to the THF core began with Keck's asymmetric allylation³⁰ of 4-pentenal (Scheme 4). The Mukaiyama's oxidative cyclization of 21^{31} thus obtained (>98% e.e.)³² proceeded without trouble to give a THF alcohol 22^{33} in good yield.¹⁹ After ozonolysis, the resulting aldehyde underwent vinylation to give a diol 7 as an epimeric mixture. Regioselective sulfonylation and silylation of two hydroxyl groups in 7 were carried out by the previously reported method.⁹ Thus, treatment of 7 with 1.1 equiv of triflic anhydride in the presence of 2,6-lutidine at -78 °C followed by addition of TBSOTf afforded **23** in one pot. Lithium enolate generated from γ -lactone **6** reacted with the triflate to give a

Scheme 3. Oxidative Cyclization of Terminal Diene Diols 9a and 9b



Table 1. Oxidative Cyclization of Diols 9a and 9b

		yield (%) ^b					
entry	conditions ^a	16	17	19	20		
1	9a, Co(modp) ₂ (20 mol %), t-BuO ₂ H (100 mol %), O ₂ , MS4A, 2-propanol, 50–52 °C	20	42	_	-		
2	9a, Co(nmp) ₂ (10 mol %), t-BuO ₂ H (10 mol %), O ₂ , 2-propanol, 50–52 °C, then MeI, 0 °C \rightarrow rt	26	57	-	-		
3	9b, Co(modp) ₂ (20 mol %), t-BuO ₂ H (100 mol %), O ₂ , MS4A, 2-propanol, 50–52 °C	-	-	50	20		
4	9b , Co(nmp) ₂ (10 mol %), <i>t</i> -BuO ₂ H (10 mol %), O ₂ , 2-propanol, 50–52 °C, then MeI, 0 °C \rightarrow rt	-	-	55	23		
$a_{\rm Co(modn)}$	= big(1 morpholinocarbamovil 4.4 dimethyl 1.2 pontanodionata) coholt(II) ¹² Co(nmp) = big	(1 1 dima	$+h_{rd} + (A)$	mathulnin	(arring)		

 $(Co(modp)_2 = bis(1-morpholinocarbamoyl-4,4-dimethyl-1,3-pentanedionate) cobalt(II).$ $(Co(mop)_2 = bis(4,4-dimethyl-1-(4-methylpiperazino)-carbamoyl-1,3-pentanedionate) cobalt(II).$ $(Bis)^{13} = bis(1-morpholinocarbamoyl-1,4-dimethyl-1-(4-methylpiperazino)-carbamoyl-1,3-pentanedionate) cobalt(II).$



coupling product 24.³⁴ Acidic hydrolysis of the TBS group followed by an allylic oxidation gave 4a in 72% yield along with the diastereomer 4b (8%).³⁵

The complete carbon skeleton of 1 was assembled by joining 3 and 4a in the presence of Grubbs' second generation catalyst³⁶ in dichloromethane to afford an enone 25 (Scheme 5). This was hydrogenated to give 26. Finally, installation of a butenolide residue and desilylation afforded structure 1. The spectroscopic and physical properties of synthetic material 1 were found to differ from those reported of the natural aromin. In particular, the natural product contained two multiplets at δ 3.59 (H-4) and 3.84 (H-7), which were observed at 4.21 and 4.36 ppm, respectively, in the ¹H NMR spectrum of the synthetic product (Table 2). These results suggested a difference in the structure around the central THF ring. Therefore, we presumed the diastereomer 31 as another possibility of aromin (Figure 2). Synthesis of 31 started from ent-21³¹ obtained by using (R)-BINOL in the asymmetric allylation (Scheme 4). By the same sequence of reactions described above, this alcohol was transformed into γ -lactone 27 including the antipodal THF ring and then enone 28.³⁷ Crossmetathesis of 28 with 3 followed by reduction of the resulting product 29 yielded 30, which was converted into 31 through a three-step sequence. Contrary to expectations, the NMR data of 31 were inconsistent with those of the natural product (Table 2). The five signals for C-4-8 of the synthetic compounds 1 and 31 deviated by 0.4-0.8 ppm compared

Scheme 5. Completion of Total Synthesis of the Proposed Structure 1 for Aromin



Table 2.	NMR	Data	$(\boldsymbol{\delta})$	for	Natural	Aromin,	Comp	oound	1,	and	Its	Diastereome	r (31
----------	-----	------	-------------------------	-----	---------	---------	------	-------	----	-----	-----	-------------	-----	----

	natural aromir	a	synthetic 1		synthetic 31		
Position	¹ H (J) (500 MHz, CDCl ₃)	$^{13}C^{c}$ (CDCl ₃)	¹ H (J) (600 MHz, CDCl ₃)	${}^{13}\text{C}^d$ (CDCl ₃)	¹ H (J) (600 MHz, CDCl ₃)	$^{13}C^d$ (CDCl ₃)	
1	-	174.2	-	173.9	-	174.0	
2	_	130.6	-	130.8	-	130.7	
3a	2.36 (m)	31.1-31.9	2.46 (m)	31.1	2.45 (m)	31.1	
3b							
4	3.59 (m)	75.6	4.21 (m)	76.4	4.20 (m)	76.4	
5a	1.22 (m)	31.1-31.9	1.62 (m)	31.6	1.58 (m)	31.6	
5b	1.56 (m)		2.08 (m)		2.06 (m)		
6a	1.22 (m)	31.1-31.9	1.53 (m)	32.0	1.52 (m)	32.0	
6b	1.56 (m)		2.14 (m)		2.13 (m)		
7	3.84 (m)	74.1	4.36 (m)	74.8	4.36 (m)	74.7	
8a	2.38	49.1	2.59	48.7	2.49	48.7	
	(dd, 15.5, 4.0)		(dd, 15.8, 5.5)		(dd, 15.8, 5.5)		
8b	2.63		2.70		2.70		
	(dd, 15.5, 9.0)		(dd, 15.8, 7.2)		(dd, 15.8, 7.2)		
9	_	209.3	-	209.3	-	209.3	
10	2.42 (t, 7.0)	43.8	2.43 (t, 7.5)	43.4	2.43 (t, 7.3)	43.4	
11	1.3-1.8	29.3-29.7	1.57 (m)	23.4	1.57 (m)	23.4	
12	1.3-1.8	25.2-25.6	1.31 (m)	29.2	1.31 (m)	29.2	
13	1.3-1.8	25.2 ^b	1.38 (m), 1.51 (m)	25.4	1.37 (m), 1.51 (m)	25.4	
14	1.39 (m)	33.5 ^b	1.39 (m)	33.2	1.39 (m)	33.2	
15	3.40 (m)	74.1 ^b	3.39 (m)	73.9	3.39 (m)	73.9	
16	3.79 (m)	82.7 ^b	3.79 (m)	82.6	3.79 (m)	82.6	
17a	1.66 (m)	28.7 ^b	1.67 (m)	28.7	1.67 (m)	28.7	
17b	1.98 (m)		1.97 (m)		1.97 (m)		
18a	1.98 (m)	28.7 ^b	1.67 (m)	28.7	1.67 (m)	28.7	
18b			1.97 (m)		1.97 (m)		
19	3.79 (m)	82.5 ^b	3.79 (m)	82.7	3.79 (m)	82.7	
20	3.40 (m)	73.7 ^b	3.39 (m)	74.0	3.39 (m)	74.0	
21	1.39 (m)	33.3 ^b	1.39 (m)	33.5	1.39 (m)	33.5	
22	1.3-1.8	25.6 ^b	1.37 (m), 1.51 (m)	25.6	1.37 (m), 1.51 (m)	25.6	
23	1.3-1.8	29.3-29.7	1.25 (m)	29.6-29.7	1.25 (m)	29.6-29.7	
24-30	1.3-1.8	23.3-31.9	1.25 (m)	29.3-31.9	1.25 (m)	29.3-31.9	
31	1.3-1.8	22.7	1.27 (m)	22.7	1.27 (m)	22.7	
32	0.88 (t, 7.0)	14.1	0.87 (t, 7.2)	14.1	0.87 (t, 6.8)	14.1	
33	7.15 (bs)	151.3	7.18 (m)	151.6	7.19 (m)	151.7	
34	4.99	77.8	5.01	77.7	5.00	77.7	
	(qddd, 6.5, 1.5, 1.5, 1.5)		(br. q. 6.8)		(br. q. 6.9)		
35	1.40 (d, 6.5)	19.1	1.40 (d, 6.8)	19.1	1.41 (q, 6.9)	19.1	
^a Reference 5	5. ^b Interchangeable within th	e same column.	^c 125 MHz. ^d 150 MHz.				

with the respective signals of the natural compound in the ¹³C

NMR spectrum.

Comparing the NMR data of natural aromin with those of acetogenins having a 4-hydroxyl group, we speculated that the natural product might possess an ether ring at the C4 position.³⁸ The fact that two signals derived from H-4 and H-7 of the natural product were observed at a high field compared to those of our synthetic samples made us reexamine the ring size of the natural product. Although ¹H NMR chemical shifts are very sensitive to neighboring substituents,



Figure 2. Structures of the diastereomer of 1 and its synthetic intermediates.

the ring size of a cyclic ether could be distinguishable by ¹³C NMR data in general. To clarify the real structure of natural aromin, we searched similar ¹³C NMR data from the literature using the CAST (CAnonical-representation of STereochemistry)/CNMR system.^{39–41} Recently one of us developed a new system CAST/CNMR Structure Elucidator⁴² and successfully revised several natural products.^{43,44} The system uses a set of ¹³C NMR chemical shifts as a query and searches partial structures with similar ¹³C NMR chemical shifts from the database developed for the CAST/CNMR Chemical Shift Predictor^{39,40,45,46} using CAST codes. By applying the CAST/CNMR chemical shift data of natural aromin, montanacin D (**32**) (Figure 3) was found as a structural candidate having a well



Figure 3. Structures of montanacin D (32) and its derivatives.

matched ¹³C NMR data (Table 3).^{47,48} This natural product was isolated from *Annona montana* by Qin et al. in 2000,³ and its structure was established by our total synthesis.^{9a}

In the reported NMR data for natural aromin, assignments of methylene groups were ambiguous, however three high-field shifted methylene carbons at 23.3 (C-6), 23.5 (C12), and 22.7 (C31) ppm of montanacin D were distinguished. Misassignment of C-6 seemed to lead to the incorrectly proposed THF structure for aromin. The ¹H NMR data well matched those reported of aromin. In order to gain further information for the identification, we prepared diacetate 33 and di-TMS ether 34 from 32. As shown in Table 3, the ¹H data of 33 in C_6D_6 were identical to those reported for aromin diacetate. MS spectral data also supported the structural revision. Aromin di-TMS ether was reported to display a fragment ion at m/z 195 corresponding to the C_1-C_8 part in the EIMS while the di-TMS derivative 35 prepared from the synthetic 1 gave a strong fragment ion at m/z 181 instead of the corresponding fragment ion (Figure 4). The fragment ion can be explained by the

cleavage between C-7 and -8 not C-8 and -9. On the other hand, the fragment ion $(m/z \ 195)$ was observed in the EIMS spectra of montanacin D bis-TMS ether 34.

These results strongly suggest that the structure of aromin should be revised to be montanacin D (32). To clarify this, direct comparison of natural aromin with our authentic sample (montanacin D) is needed. Since aromin and aromicin differ only by the length of the carbon chain, reinvestigation of the proposed structure for aromicin is necessary.⁴⁹

Antitumor activities of synthetic samples 1 and 31 were evaluated, and their data are summarized in Table 4. Both compounds showed significant cytotoxicities against the six human solid tumor cell lines tested *in vitro*. The level of activity was revealed to be relatively strong compared to those of THP acetogenins^{9b} such as 32 but not to be comparable to those expected for the usual nonadjacent bis-THF acetogenins.

In summary, the usefulness of a cross-olefin metathesis and a Co-mediated oxidative cyclization of a bis-homoallyl alcohol was demonstrated by the total synthesis of the proposed structure for aromin. Our synthetic studies coupled with a structure search system using the CAST/CNMR Structure Elucidator revealed that aromin possessing an unusual bis-THF ring structure should be revised to a THP acetogenin, montanacin D (32). In addition, these results obtained here suggest that the structures of related natural products such as aromicin and aromin-A^{4,49} should be reinvestigated.

EXPERIMENTAL SECTION

General Procedures. All reactions were carried out under an argon atmosphere, unless otherwise noted. IR spectra were recorded by the ATR method. The NMR spectra were recorded at 500 or 600 MHz for ¹H and 125 or 150 MHz for ¹³C. Chemical shifts are reported in ppm downfield from tetramethylsilane with the solvent resonance as the internal standard ($\delta_{\rm H}$ 7.26 ppm or $\delta_{\rm C}$ 77.0 ppm). High-resolution mass spectra (HRMS) were acquired in the electron ionization mode (EI) or the field ionization mode (FI) using a gas chromatography time-of-flight mass spectrometer or electrospray ionization (ESI) hybrid quadrupole/time-of-flight tandem mass spectrometer. The solvent extracts were dried with magnesium sulfate, and the solutions were evaporated under diminished pressure at 35–40 °C.

(4R,5R)-4-(But-3-en-1-yl)-2,2-dimethyl-5-(pent-4-en-1-yl)-1,3-dioxolane (11). To a stirred solution of 10 (1.0 g, 4.99 mmol) and N,N-diisopropylethylamine (2.62 mL, 15.0 mmol) in dichloromethane (6.0 mL) was added dropwise a solution of trifluoromethanesulfonic anhydride (1.26 mL, 7.48 mmol) in dichloromethane (84 mL) at -40 °C, and the mixture was stirred at the same temperature for 30 min. After addition of ice-water, the resulting mixture was stirred vigorously for 10 min and then extracted with dichloromethane. The combined organic layers were washed successively with water, saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was passed through a short column of silica gel (*n*-hexane–ethyl acetate = $50:1 \rightarrow 10:1$) to give a syrup (1.55 g). This compound was employed to the next step without further purification. To a stirred suspension of CuBr (273 mg, 1.90 mmol) in ether (15 mL) was added a 1.0 M solution of allylmagnesium bromide in ether (15 mL, 15.0 mmol) at 0 °C, and the mixture was stirred at the same temperature for 20 min. A solution of the above-mentioned syrup (1.55 g) in ether (8 mL) was added dropwise at 0 °C, and the mixture was stirred at 0 °C \rightarrow rt for 14 h. After addition of saturated aqueous NH₄Cl, the mixture was extracted with dichloromethane-ethyl acetate (1:1). The combined organic layers were washed successively with water and brine, dried, and concentrated. The residue was chromatographed on silica gel (nhexane-ether = $50:1 \rightarrow 10:1$) to give 11 (1.0 g, 89% from 10) as a light-yellow liquid; $[\alpha]_D^{25}$ +30.3 (c 0.44, CHCl₃); IR (ZnSe) 3077, 2984, 2932, 1641, 1235, 1087, 908, 875 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.86–5.75 (2H, m), 5.06–4.94 (4H, m), 3.63–3.59 (2H,

Table 3. NMR Data (δ) for Montanacin D (32), Its Diacetate 33, and Natural Aromin-15,20 Diacetates

	montanacin D $(32)^{a}$		33		aromin-15,20 diacetates ^b		
Position	¹ H (J) (600 MHz, CDCl ₃)	$^{13}C^{c}$ (CDCl ₃)	¹ H (J) (600 MHz, C ₆ D ₆)	Position	¹ H (J) (500 MHz, C ₆ D ₆)		
1	-	174.1	-	1	_		
2	-	130.6	-	2	-		
3a	2.35 (m)	31.8	2.23 (m)	3a	2.22 (dddd, 16.0, 3.5, 1.5, 1.5)		
3b	2.35 (m)		2.32 (m)	3b	2.33 (dddd, 16.0, 9.0, 1.5, 1.5)		
4	3.58 (m)	75.6	3.33 (m)	4	3.34 (m)		
5a	1.25 (m)	31.1	0.98 (m)	5a	0.98 (m)		
5b	1.58 (m)		1.19 (m)	5b	1.17 (m)		
6a	1.58 (m)	23.3	1.48 (m)	6a	0.93 (m)		
6b	1.84 (m)		1.48 (m)	6b	1.20 (m)		
7a	1.23 (m)	31.1	0.96 (m)	7	3.66 (m)		
7b	1.58 (m)		1.19 (m)	8a	1.93 (dd, 15.5, 3.5)		
8	3.82 (m)	74.1	3.66 (m)	8b	2.37 (dd, 15.5, 9.0)		
9a	2.37 (dd, 15.9, 8.8)	49.1	1.92 (dd, 15.8, 3.8)	9	-		
9b	2.62 (dd, 15.9, 3.9)		2.34 (dd, 15.8, 9.2)				
10	_	209.3	_	10	2.08 (t, 7.0)		
11	2.41 (t, 7.2)	43.8	2.08 (m)	11	0.94-1.76		
12	1.52 (m), 1.58 (m)	23.5	1.48 (m), 1.56 (m)	12	0.94-1.76		
13	1.36 (m), 1.50 (m)	25.2	1.32 (m)	13	0.94-1.76		
14	1.39 (m)	33.2	1.51 (m), 1.59 (m)	14	1.5 ^d		
15	3.38 (m)	73.7	5.02 (m)	15	5.05 (m)		
16	3.79 (m)	82.5	3.97 (m)	16	3.97 (m)		
17a	1.68 (m)	28.7	1.39 (m)	17a	1.41 (m)		
17b	1.98 (m)		1.65 (m)	17b	1.65 (m)		
18a	1.68 (m)	28.7	1.39 (m)	18a	1.41 (m)		
18b	1.98 (m)		1.65 (m)	18b	1.65 (m)		
19	3.79 (m)	82.7	3.98 (m)	19	3.97 (m)		
20	3.40 (m)	74.0	5.06 (m)	20	5.05 (m)		
21	1.39 (m)	33.5	1.54 (m), 1.63 (m)	21	1.7 ^d		
22	1.36 (m), 1.50 (m)	25.6	1.39 (m)	22	0.97-1.76		
23	1.25 (m)	29.6-29.7	1.30 (m)	23	0.97-1.76		
24-30	1.25 (m)	29.6-29.7	1.30 (m)	24-30	0.97-1.76		
31	1.28 (m)	22.7	1.30 (m)	31	0.97-1.76		
32	0.87 (t, 6.9)	14.1	0.91 (t, 7.0)	32	0.91 (t, 7.0)		
33	7.14 (m)	151.3	6.65 (m)	33	6.65 (ddd, 1.5, 1.5, 1.5)		
34	4.98 (brq, 6.6)	77.8	4.51 (brq, 6.5)	34	4.51 (qddd, 6.5, 1.5, 1.5, 1.5)		
35	1.40 (d, 6.6)	19.1	1.02 (d, 6.5)	35	1.02 (d, 6.5)		
15-OAc			1.82 (s)	15-OAc	1.82 $(s)^d$		
20-OAc			1.83 (s)	20-OAc	$1.83 (s)^d$		

^aReference 9a. ^bReference 5. ^c150 MHz. ^dInterchangeable within the same column.



Figure 4. EIMS data of aromin di-TMS ether (35) and montanacin D di-TMS ether (34).

Table 4. ED₅₀ (μ g/mL) Values of 1 and 31 against Six Human Solid Tumor Cell Lines^a

compd	A-549	MCF-7	HT-29	A-498	PC-3	PACA-2
1	2.1	1.1×10^{-1}	2.3	7.3×10^{-1}	5.8×10^{-2}	3.5×10^{-1}
31	1.2	8.6×10^{-2}	1.9	6.0×10^{-1}	3.8×10^{-2}	1.1×10^{-1}
Adr ^b	8.4×10^{-2}	6.6×10^{-2}	1.8×10^{-2}	1.9×10^{-2}	8.4×10^{-2}	9.0×10^{-3}

^{*a*}A-549 (lung carcinoma), MCF-7 (breast carcinoma), HT-29 (colon adenocarcinoma), A-498 (renal carcinoma), PC-3 (prostate adenocarcinoma), PACA (pancreas carcinoma). ^{*b*}Adriamycin was used for the standard positive control.

m), 2.24 (1H, m), 2.14 (1H, m), 2.11–2.05 (2H, m), 1.64–1.42 (6H, m), 1.37 (6H, s); ¹³C NMR (125 MHz, CDCl₃): δ 138.4, 138.0, 114.8, 114.7, 107.9, 80.7, 80.2, 33.7, 32.2, 32.1, 30.2, 27.3, 25.3; HRMS (EI) calcd for C₁₃H₂₁O₂ [M – Me]⁺ 209.1542, found 209.1547.

(5*R*,6*R*)-Undeca-1,10-diene-5,6-diol (8). To a stirred solution of 11 (224 g, 1.0 mmol) in methanol (10 mL) was added a 10% HCl solution in methanol (0.3 mL). The mixture was stirred at 70 °C for 4.5 h, cooled, concentrated *in vacuo*, and then coevaporated with ethanol-benzene (×5). The residue was chromatographed on silica gel (*n*-hexane–ethyl acetate = 10:1 → 4:1) to give 8 (170 mg, 92%) as a syrup; $[\alpha]_D^{26}$ +27.8 (*c* 0.78, CHCl₃); IR (ZnSe) 3358, 3076, 2916, 1640, 1070, 992, 906 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.86–5.75 (2H, m), 5.06–4.93 (4H, m), 3.39 (2H, m), 2.70 (2H, brs), 2.26 (1H, m), 2.14 (1H, m), 2.11–2.03 (2H, m), 1.61–1.39 (6H, m); ¹³ C NMR (125 MHz, CDCl₃): δ 138.5, 138.3, 115.0, 114.7, 74.3, 73.9, 33.6, 32.8, 32.6, 29.9, 24.8; HRMS (ESI) calcd for C₁₁H ₂₀O₂ Na[M + Na] ⁺ 207.1361, found 207.1366.

(R)-1-((2R,5R)-5-(Hydroxymethyl)tetrahydrofuran-2-yl)hex-5-en-1-ol (5). To a stirred solution of 8 (75 mg, 0.41 mmol) and Co(nmp)₂ (23 mg, 0.04 mmol) in 2-propanol (4.0 mL) was added a 5.0–6.0 M solution of *tert*-butyl hydroperoxide in decane (10 μ L), and the mixture was stirred at 50-52 °C for 8.5 h under an oxygen atmosphere and then cooled to rt. After addition of iodomethane (25 μ L) at 0 °C, the resulting mixture was stirred at 0 °C \rightarrow rt for 13 h, concentrated, diluted with water, and then extracted with dichloromethane. The combined organic layers were washed with brine, dried, and concentrated. The residue was chromatographed on silica gel (nhexane-ethyl acetate = $10:1 \rightarrow 4:1 \rightarrow 1:1$) to give 5 (63.6 mg, 78%) as a syrup; $[\alpha]_D^{26}$ +5.7 (c 0.34, CHCl₃); IR (ZnSe) 3398, 3076, 2915, 1639, 1235, 1085, 878 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.80 (1H, ddt, J = 17.2, 10.3, 6.7 Hz), 5.00 (1H, ddt, J = 17.2, 2.0, 1.5 Hz), 4.94 (1H, brd, J = 10.3, 2.0, 1.0 Hz), 4.10 (1H, m), 3.82 (1H, dt, J = 7.9, 6.6 Hz), 3.67 (1H, brd, J = 11.1 Hz), 3.50 (1H, brdd, J = 11.1, 5.1 Hz), 3.42 (1H, m), 2.73 (1H, brs), 2.47 (1H, brs), 2.10-1.95 (4H, m), 1.73-1.62 (3H, m), 1.51-1.40 (3H, m); ¹³ C NMR (125 MHz, CDCl₃): δ 138.6, 114.6, 82.9, 79.7, 74.0, 64.7, 33.7, 32.6, 28.5, 27.8, 24.8; HRMS (ESI) calcd for $C_{11}H_{20}O_3Na [M + Na]^+$ 223.1310, found 223.1311.

((2R,5R)-5-((R)-1-((tert-Butyldimethylsilyl)oxy)hex-5-en-1-yl)tetrahydrofuran-2-yl)methanol (12). To a stirred solution of 5 (41 mg, 0.21 mmol) and 2,6-lutidine (0.12 mL, 1.03 mmol) in dichloromethane (1.3 mL) was added tert-butyldimethylsilyl trifluoromethanesulfonate (0.12 mL, 0.52 mmol) at 0 °C, and the mixture was stirred at 0 $^{\circ}C \rightarrow$ rt for 3 h. After addition of ice-water, the resulting mixture was stirred vigorously for 40 min and then extracted with ether. The combined organic layers were washed successively with cold aqueous HCl, water, saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was passed through a short column of silica gel (*n*-hexane-ether = $100:1 \rightarrow 10:1$) to give a syrup (85 mg) which was dissolved in tetrahydrofuran (1.0 mL). To the solution was added a 1.0 M solution of tetrabutylammonium fluoride in tetrahydrofuran (0.15 mL) at -78 °C, and the mixture was stirred at -78 °C \rightarrow rt for 23 h, diluted with ethyl acetate, washed with brine, dried, and concentrated. The residue was chromatographed on silica gel (*n*-hexane-ethyl acetate = $10:1 \rightarrow 4:1$) to give 12 (48.1 mg, 75%) as a syrup; $[\alpha]_D^{27}$ +7.4 (c 0.80, CHCl₃); IR (ZnSe) 3392, 3077, 2927, 2855, 1640, 1250, 1076, 878, 831 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.80 (1H, ddt, J = 17.1, 10.3, 6.6 Hz), 5.00 (1H, ddt, J = 17.3, 2.2, 1.7 Hz), 4.94 (1H, ddt, J = 10.3, 2.2, 1.2 Hz), 4.08 (1H, m), 3.91 (1H, dt, J = 7.8, 6.1 Hz), 3.64 (1H, ddd, J = 11.7, 5.9, 3.2 Hz), 3.58 (1H, m), 3.48 (1H, dt, J = 11.7, 5.4 Hz), 2.10 (1H, brs), 2.10–1.98 (2H, m), 1.97-1.88 (2H, m), 1.74-1.61 (2H, m), 1.55-1.32 (4H, m), 0.88 (9H, s), 0.07 (3H, s), 0.06 (3H, s); ¹³C NMR (150 MHz, CDCl₂): δ 138.8, 114.5, 82.0, 79.4, 74.8, 64.9, 33.8, 32.4, 27.8, 27.7, 26.0, 24.9, 18.3, –4.2, –4.6; HRMS (ESI) calcd for $C_{17}H_{34}O_3SiNa\ [M$ + $Na]^4$ 337.2175, found 337.2179.

(S)-1-((2R,5R)-5-((R)-1-((tert-Butyldimethylsilyl)oxy)hex-5-en-1-yl)tetrahydrofuran-2-yl)tridecan-1-ol (13) and (R)-1-((2R,5R)-5-((R)-1-((tert-Butyldimethylsilyl)oxy)hex-5-en-1-yl)tetrahydrofuran-2-yl)tridecan-1-ol (14). To a stirred solution of oxalyl chloride (85 μ L, 0.97 mmol) in dichloromethane (2.0 mL) was added dropwise a solution of DMSO (0.15 mL, 1.94 mmol) in dichloromethane (0.5 mL) at -70 °C, and the mixture was stirred at -70 °C for 1 h. A solution of 12 (68 mg, 0.22 mmol) in dichloromethane (0.6 mL) was added dropwise, and the mixture was stirred at the same temperature for 1 h. Triethylamine (0.30 mL, 2.16 mmol) was added, and the resulting mixture was gradually warmed to 0 °C with stirring and then poured into ice-water. The resulting mixture was extracted with ether. The combined organic layers were washed successively with cold aqueous HCl, water, saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was coevaporated with benzene $(\times 5)$ to give a syrup (72) mg) which was dissolved in ether (1.5 mL). To this stirred solution was added dropwise a 1.0 M solution of dodecylmagnesium bromide (1.76 mL, 1.76 mmol) in ether at -78 °C, and the mixture was stirred at the same temperature for 4.5 h. Saturated aqueous NH₄Cl was added with vigorous stirring, and then the resulting mixture was extracted with ether. The combined organic layers were washed successively with water and brine, dried, and concentrated. The residue was passed through a short column of silica gel (n-hexane-ethyl acetate = $10:1 \rightarrow 4:1$) to give a syrup (104 mg), which was purified by preparative TLC (*n*-hexane-ethyl acetate = 10:1, five developments)

to afford 13 (19 mg, 18%) and 14 (76 mg, 73%). 13. Syrup; $[\alpha]_D^{27}$ +14.3 (*c* 0.30, CHCl₃); IR (ZnSe) 3464, 3073, 2923, 2853, 1640, 1467, 1236, 1087, 883, 834, 774 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.80 (1H, ddt, *J* = 17.0, 10.0, 6.6 Hz), 5.00 (1H, ddt, *J* = 17.0, 2.0, 1.7 Hz), 4.94 (1H, ddt, *J* = 10.0, 2.0, 1.2 Hz), 3.91 (1H, dt, *J* = 8.6, 6.3 Hz), 3.84 (1H, m), 3.79 (1H, m), 3.55 (1H, m), 2.08–2.02 (3H, m), 1.92 (1H, m), 1.83–1.78 (2H, m), 1.59 (1H, m), 1.61–1.25 (26H, m), 0.884 (9H, s), 0.876 (3H, t, *J* = 7.4 Hz), 0.07 (3H, s), 0.05 (3H, s); ¹³C NMR (125 MHz, CDCl₃): δ 138.8, 114.4, 82.5, 82.1, 75.2, 71.6, 33.8, 32.5, 32.4, 31.9, 29.73, 29.66, 29.65, 29.63, 29.59, 29.56, 29.3, 28.0, 26.00, 25.97, 25.1, 24.8, 22.7, 18.3, 14.1, -4.1, -4.6; HRMS (ESI) calcd for C₂₉H₅₈O₃SiNa [M + Na]⁺ 505.4053, found 505.4061.

14. Syrup; $[\alpha]_D^{27}$ +14.7 (*c* 0.63, CHCl₃); IR (ZnSe) 3574, 3069, 2923, 2853, 1640, 1465, 1236, 1088, 879, 834, 774 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.79 (1H, ddt, *J* = 17.2, 10.3, 6.6 Hz), 4.99 (1H, ddt, *J* = 17.2, 2.0, 1.6 Hz), 4.94 (1H, ddt, *J* = 10.3, 2.0, 1.2 Hz), 3.84 (1H, dt, *J* = 8.0, 6.4 Hz), 3.76 (1H, dt, *J* = 7.8, 6.6 Hz), 3.56 (1H, m), 3.37 (1H, m), 2.39 (1H, d, *J* = 3.9 Hz), 2.07–1.99 (2H, m), 1.95–1.89 (2H, m), 1.70–1.55 (2H, m), 1.52–1.24 (26H, m), 0.88 (9H, s), 0.87 (3H, t, *J* = 7.1 Hz), 0.07 (3H, s), 0.05 (3H, s); ¹³C NMR (125 MHz, CDCl₃): δ 138.7, 114.5, 82.4, 82.2, 75.1, 74.1, 33.8, 33.4, 32.5, 31.9, 29.71, 29.66, 29.65, 29.63, 29.60, 29.57, 29.3, 28.55, 28.45, 25.9, 25.6, 24.6, 22.7, 18.3, 14.1, -4.1, -4.6; HRMS (ESI) calcd for C₂₉H₅₈O₃SiNa [M + Na]⁺ 505.4053, found 505.4052.

tert-Butyl(((R)-1-((2R,5R)-5-((R)-1-((tert-butyldimethylsilyl)oxy)hex-5-en-1-yl)tetrahydrofuran-2-yl)tridecyl)oxy)dimethylsilane (3). To a stirred solution of 14 (35 mg, 72 μ mol) and 2,6lutidine (21 μ L, 0.18 mmol) in dichloromethane (0.8 mL) was added *tert*-butyldimethylsilyl trifluoromethanesulfonate (18 μ L, 80 μ mol) at 0 °C, and the mixture was stirred at 0 °C for 2 h. After addition of icewater, the resulting mixture was stirred vigorously for 20 min and then extracted with ether. The combined organic layers were washed successively with cold aqueous HCl, water, saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was chromatographed on silica gel (*n*-hexane–ether = $200:1 \rightarrow 100:1$) to give 3 (41 mg, 96%) as a syrup; $[\alpha]_D^{25}$ +20.2 (c 0.88, CHCl₃); IR (ZnSe) 2924, 2854, 1636, 1471, 1235, 1086, 873, 833, 773 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.81 (1H, ddd, J = 17.1, 10.0, 6.6 Hz), 5.00 (1H, ddt, J = 17.1, 2.0, 1.6 Hz), 4.94 (1H, ddt, J = 10.0, 2.0, 1.0 Hz), 3.90 (1H, m), 3.58 (1H, m), 2.04 (1H, m), 1.83 (1H, m), 1.68 (1H, m), 1.51 (1H, m), 1.43-1.22 (28H, m), 0.88 (18H, s), 0.87 (3H, t, I = 7.4 Hz), 0.058 (3H, s), 0.054 (3H, s), 0.049 (3H, s), 0.047 (3H, s); ¹³C NMR (125 MHz, CDCl₃): δ 138.9, 114.3, 81.7, 74.7, 74.6, 33.9, 32.6, 32.0, 31.9, 29.8, 29.69, 29.66, 29.65, 29.63, 29.62, 29.4, 27.2, 25.95, 25.87, 25.2, 22.7, 18.2, 14.1, -4.3, -4.55. -4.57; HRMS (ESI) calcd for $C_{35}H_{72}O_3Si_2Na [M + Na]^+$ 619.4918, found 619.4918.

(35,5*R*)-Nona-1,8-diene-3,5-diol (9a). To a stirred solution of 37 (563 mg, 2.30 mmol) in acetic acid (10 mL) was added water (1.0 mL). The mixture was stirred at 50 °C for 3.5 h, cooled, concentrated, and then coevaporated with toluene (×3). The residue was chromatographed on silica gel (*n*-hexane–ethyl acetate = 3:1 → 2:1) to give 9a (359 mg, quant.) as a syrup; $[\alpha]_{\rm D}^{24}$ +1.2 (*c* 0.91, CHCl₃); IR (ZnSe) 3345, 3078, 2980, 2936, 1641, 1416, 1078, 991, 910, 825 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.84 (1H, ddd, *J* = 17.1, 10.5, 5.3 Hz), 5.76 (1H, ddt, *J* = 17.1, 10.0, 6.7 Hz), 5.21 (1H, ddd, *J* = 17.1, 1.5, 1.5 Hz), 5.05 (1H, ddd, *J* = 10.5, 1.5, 1.2 Hz), 4.98 (1H, ddt, *J* = 17.1, 1.8, 1.6 Hz), 4.91 (1H, brdd, *J* = 10.0, 1.8 Hz), 4.39 (1H, m), 3.88 (1H, brs), 3.86 (1H, m), 3.72 (1H, brs), 2.12 (1H, m), 2.05 (1H, m), 1.65–1.52 (3H, m), 1.47 (1H, m); ¹³C NMR (125 MHz, CDCl₃): δ 140.5, 138.2, 114.6, 114.1, 69.9, 68.2, 42.2, 36.3, 29.8; HRMS (FI) calcd for C₉H₁₆O₂ [M]⁺ 156.1150, found 156.1161.

(2R,3S,5R)-5-(But-3-en-1-yl)-3-acetoxy-2-(acetoxymethyl)tetrahydrofuran (16) and (2R,5R)-2-((S)-2-acetoxybut-3-en-1yl)-5-(acetoxymethyl)tetrahydrofuran (17). (i) To a stirred solution of 9a (110 mg, 0.70 mmol) and Co(nmp)₂ (40 mg, 70 μ mol) in 2-propanol (6.9 mL) was added a 5.0–6.0 M solution of tertbutyl hydroperoxide in decane (14 μ L), and the mixture was stirred at 50-52 °C for 7 h under an oxygen atmosphere and then cooled to rt. After addition of iodomethane $(80 \ \mu L)$ at 0 °C, the resulting mixture was stirred at 0 °C \rightarrow rt for 13 h, concentrated, diluted with water, and then extracted with dichloromethane. The combined organic layers were washed with brine, dried, and concentrated. The residue was dissolved in pyridine (1.0 mL). To the solution were added acetic anhydride (0.5 mL) and N,N-dimethylaminopyridine (7.0 mg), and the mixture was stirred at rt for 14 h. After addition of ice-water, the resulting mixture was stirred vigorously for 3 h and then extracted with ether. The combined organic layers were washed successively with cold aqueous HCl, water, saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was chromatographed on silica gel (dichloromethane-ethyl acetate = $40:1 \rightarrow 30:1 \rightarrow 20:1 \rightarrow$ 10:1) to give 17 (102 mg, 57%) and 16 (46 mg, 26%).

(ii) To a stirred suspension of 9a (103 mg, 0.66 mmol), Co(modp)₂ (71.1 mg, 0.13 mmol), and MS4A (0.31 g) in 2-propanol (6.2 mL) was added a 5.0–6.0 M solution of *tert*-butyl hydroperoxide in decane (0.13 mL), and the mixture was stirred at 50–52 °C for 4 h under an oxygen atmosphere, cooled to rt, and then filtered through a pad of Celite. After addition of aqueous saturated $Na_2S_2O_3$ to the filtrate, the resulting mixture was stirred at rt for 1 h, concentrated, diluted with ethyl acetate, washed with brine, dried, and concentrated. The residue was submitted to acetylation as described above to give 17 (70.7 mg, 42%) and 16 (33.3 mg, 20%).

16. Syrup; $[\alpha]_D^{22}$ +30.0 (*c* 0.84, CHCl₃); IR (ZnSe) 3080, 2934, 1738, 1640, 1438, 1366, 1229, 1041, 906 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.82 (1H, ddt, *J* = 17.1, 10.3, 6.6 Hz), 5.09 (1H, ddd, *J* = 7.3, 4.6, 3.4 Hz), 5.03 (1H, ddt, *J* = 17.1, 1.7, 1.7 Hz), 4.97 (1H, ddt, *J* = 10.3, 1.7, 1.2 Hz), 4.19–4.10 (4H, m), 2.47 (1H, dt, *J* = 13.5, 7.1 Hz), 2.16–2.10 (2H, m), 2.09 (3H, s), 2.07 (3H, s), 1.79 (1H, m), 1.70 (1H, ddd, *J* = 13.5, 6.8, 4.6 Hz), 1.59 (1H, m); ¹³C NMR (125 MHz, CDCl₃): δ 170.7, 170.6, 137.9, 114.8, 80.5, 78.2, 75.7, 64.0, 37.5, 34.8, 30.1, 21.0, 20.8; HRMS (FI) calcd for C₁₃H₂₀O₅ [M]⁺ 256.1311, found 256.1327.

17. Syrup; $[a]_D^{22} - 27.8$ (*c* 1.03, CHCl₃); IR (ZnSe) 3090, 2938, 1737, 1647, 1435, 1370, 1232, 1086, 1022, 884 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.76 (1H, ddd, *J* = 17.3, 10.8, 6.4 Hz), 5.30 (1H, brdd, *J* = 6.4, 5.2 Hz), 5.21 (1H, ddd, *J* = 17.3, 1.2, 1.2 Hz), 5.12 (1H, ddd, *J* = 10.8, 1.2, 1.0 Hz), 4.17 (1H, m), 4.01 (1H, dd, *J* = 11.5, 3.9 Hz), 3.97 (1H, dd, *J* = 11.5, 6.8 Hz), 3.96 (1H, m), 2.06-2.00 (2H, m), 2.05 (3H, s), 2.03 (3H, s), 1.85 (1H, ddd, *J* = 13.9, 7.2, 5.3 Hz), 1.75 (1H, ddd, *J* = 13.9, 8.6, 5.9 Hz), 1.61-1.53 (2H, m); ¹³C NMR (125 MHz, CDCl₃): δ 170.9, 170.0, 136.4, 116.4, 75.8, 75.7, 72.2, 66.4, 39.9, 31.8, 28.2, 21.1, 20.8; HRMS (FI) calcd for C₁₃H₂₀O₅ [M]⁺ 256.1311, found 256.1295.

(S)-1-((2*R*,5*R*)-5-(Hydroxymethyl)tetrahydrofuran-2-yl)but-3en-2-ol (7a). To a stirred solution of 17 (102 mg, 0.39 mmol) in methanol-dichloromethane (10:1, 1.1 mL) was added a 1.0 M solution of sodium methoxide in methanol (50 μ L, 0.05 mmol), and the mixture was stirred at rt for 4 h and made neutral with Dowex-50W X-8 (H⁺) resin. The mixture was filtered, and the filtrate was evaporated. The residue was chromatographed on silica gel (*n*hexane–ethyl acetate = 2:1 \rightarrow 1:1 \rightarrow 1:5) to give 7a (57.7 mg, 93%) as a syrup; $[\alpha]_D^{25}$ –10.6 (*c* 1.00, CHCl₃); IR (ZnSe) 3346, 2932, 2869, 1646, 1037, 991, 917 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.87 (1H, ddd, *J* = 17.2, 10.3, 5.4 Hz,), 5.25 (1H, dt, *J* = 17.2, 1.5 Hz), 5.07 (1H, dt, *J* = 10.3, 1.5 Hz), 4.34 (1H, brs), 4.20 (1H, m), 4.11(1H, m), 3.60 (1H, dd, *J* = 11.7, 1.9 Hz), 3.52 (1H, brs), 3.46 (1H, dd, *J* = 11.7, 5.9 Hz), 3.08 (1H, brs), 2.03 (1H, m), 1.95 (1H, m), 1.74–1.63 (3H, m), 1.58 (1H, m); ¹³C NMR (125 MHz, CDCl₃): δ 140.8, 114.1, 79.4, 76.1, 70.0, 64.6, 41.6, 32.2, 27.1; HRMS (FI) calcd for C₉H₁₇O₃ [M + H] ⁺ 173.1178, found 173.1177.

(35,55)-Nona-1,8-diene-3,5-diol (9b). Compound 38 (488 mg, 1.99 mmol) in acetic acid-water (10:1; 11 mL) was heated at 50-55 °C with stirring for 9.0 h. After being cooled to rt, the resulting mixture was concentrated and then coevaporated with toluene $(\times 3)$. The residue was chromatographed on silica gel (n-hexane-ethyl acetate = $3:1 \rightarrow 2:1$) to give 9b (210 mg, 67%) and recovered 38 (200 mg). The latter was converted into 9b (84 mg) by treating with acetic acid-water (10:1; 5.5 mL) at 50-55 °C for 6.0 h, followed by chromatography on silica gel (n-hexane-ethyl acetate = $3:1 \rightarrow 2:1$). The total amount of 9b was 294 mg (94%); $[\alpha]_{D}^{22}$ +0.5 (c 2.01, CHCl₃); IR (ZnSe) 3329, 3078, 2977, 2936, 1641, 1421, 1312, 1134, 1078, 990, 909, 846 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.87 (1H, ddd, J = 17.4, 10.5, 5.9 Hz), 5.83 (1H, ddt, J = 17.2, 10.2, 6.7 Hz), 5.24 (1H, ddd, J = 17.4, 1.5, 1.2 Hz), 5.09 (1H, ddd, J = 10.5, 1.4, 1.2 Hz), 5.04 (1H, ddt, J = 17.2, 2.0, 1.5 Hz), 4.97 (1H, ddt, J = 10.2, 2.0, 1.2 Hz), 4.36 (1H, m), 3.91 (1H, m), 2.94 (2H, brs), 2.18-2.12 (2H, m), 1.67–1.42 (4H, m); ¹³C NMR (125 MHz, CDCl₃): δ 140.6, 138.3, 114.9, 114.5, 73.7, 71.9, 42.8, 37.0, 29.7; HRMS (FI) calcd for $C_9H_{16}O_2$ [M]⁺ 156.1150, found 156.1171.

(25,55)-2-((S)-2-Acetoxybut-3-en-1-yl)-5-(acetoxymethyl)tetrahydrofuran (19) and (25,35,55)-5-(But-3-en-1-yl)-3-acetoxy-2-(acetoxymethyl)tetrahydrofuran (20). (i) Treatment of 9b (144 mg, 0.922 mmol) with $Co(nmp)_2$ (52.1 mg, 0.09 mmol) and *tert*-butyl hydroperoxide in decane (18 μ L) as described for preparation of 16 and 17 gave 19 (129.8 mg, 55%) and 20 (55.1 mg, 23%).

(ii) Treatment of **9b** (133 mg, 0.85 mmol) with $Co(modp)_2$ (92 mg, 0.17 mmol) and *tert*-butyl hydroperoxide in decane (0.17 mL) as described for preparation of **16** and **17** gave **19** (109 mg, 50%) and **20** (44.1 mg, 20%).

19. Syrup; $[\alpha]_D^{24}$ +10.8 (*c* 1.50, CHCl₃); IR (ZnSe) 3087, 2923, 1736, 1664, 1436, 1370, 1233, 1086, 1039, 887 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.80 (1H, ddd, *J* = 17.1, 10.5, 6.4 Hz), 5.38 (1H, dt, *J* = 7.4, 6.3 Hz), 5.27 (1H, ddd, *J* = 17.1, 1.2, 1.2 Hz), 5.18 (1H, ddd, *J* = 10.5, 1.2, 1.2 Hz), 4.23 (1H, m), 4.10 (1H, dd, *J* = 11.5, 3.5 Hz), 4.00 (1H, m), 3.97 (1H, dd, *J* = 11.5, 7.1 Hz), 2.10–2.01 (3H, m), 2.09 (3H, s), 2.06 (3H, s), 1.71 (1H, dt, *J* = 13.9, 6.1 Hz), 1.64–1.52 (2H, m); ¹³C NMR (125 MHz, CDCl₃): δ 171.1, 170.2, 136.1, 116.9, 76.04, 75.97, 72.5, 66.6, 39.9, 32.0, 28.2, 21.3, 21.0; HRMS (FI) calcd for C₁₃H₂₀O₅ [M]⁺ 256.1311, found 256.1325.

20. Syrup; $[\alpha]_D^{23}$ +14.9 (*c* 0.75, CHCl₃); IR (ZnSe) 3076, 2973, 2926, 1738, 1640, 1437, 1372, 1228, 1085, 1043, 889 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.82 (1H, ddt, *J* = 17.1, 10.3, 6.6 Hz), 5.43 (1H, brt, *J* = 4.2 Hz), 5.03 (1H, ddt, *J* = 17.1, 1.7, 1.7 Hz), 4.97 (1H, ddt, *J* = 10.3, 1.7, 1.3 Hz), 4.26 (1H, dt, *J* = 7.3, 4.2 Hz), 4.21 (1H, dd, *J* = 11.5, 5.6 Hz), 4.20 (1H, m), 4.15 (1H, dd, *J* = 11.5, 7.4 Hz), 2.20–2.06 (3H, m), 2.07 (6H, s), 1.84 (1H, ddd, *J* = 13.5, 9.8, 5.4 Hz), 1.76 (1H, ddt, *J* = 13.5, 9.4, 6.9 Hz), 1.57 (1H, m); ¹³C NMR (125 MHz, CDCl₃): δ 170.8, 170.2, 137.9, 114.9, 77.70, 77.67, 74.7, 62.8, 39.1, 34.7, 30.0, 20.99, 20.91; HRMS (FI) calcd for C₁₃H₂₀O₅ [M]⁺ 256.1311, found 256.1329.

(S)-1-((25,55)-5-(Hydroxymethyl)tetrahydrofuran-2-yl)but-3en-2-ol (7b). Treatment of 19 (130 mg, 0.51 mmol) with a 1.0 M solution of sodium methoxide in methanol (50 μ L, 0.05 mmol) as described for preparation of 7a yielded 7b (78 mg, 98%) as a syrup; $[\alpha]_{\rm D}^{22}$ +12.5 (*c* 1.01, CHCl₃); IR (ZnSe) 3355, 2926, 2854, 1647, 1420, 1376, 1036, 991, 918 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.82 (1H, ddd, J = 16.2, 10.3, 5.6 Hz), 5.22 (1H, dt, J = 16.2, 1.5 Hz), 5.05 (1H, dt, J = 10.3, 1.5 Hz), 4.28 (1H, m), 4.16 (1H, m), 4.13 (1H, m), 3.76 (1H, brs), 3.60 (1H, dd, J = 11.7, 3.2 Hz), 3.47 (1H, dd, J = 11.7, 5.9 Hz), 2.80 (1H, brs), 2.09 (1H, m), 1.94 (1H, m), 1.70–1.64 (3H, m), 1.57 (1H, m); ¹³C NMR (125 MHz, CDCl₃): δ 140.4, 114.2, 79.6, 79.0, 72.4, 64.6, 42.4, 32.7, 26.9; HRMS (FI) calcd for C₉H₁₇O₃ [M + H]⁺ 173.1178, found 173.1172.

(R)-Octa-1,7-dien-4-ol (21). To a stirred suspension of (S)-(+)-l,l'-bi-2-naphthol (1.74 g, 6.1 mmol) and MS4A (14.7 g) in dichloromethane (62 mL) was added (i-PrO)₄Ti (1.79 mL, 6.1 mmol), and the mixture was heated at reflux for 1 h and then cooled to rt. A solution of 4-pentenal (2.56 g, 30.4 mmol) in dichloromethane (10 mL) was added. After being stirred for 15 min, the contents were cooled to -78 °C, and allyltributyltin (11.1 g, 33.4 mmol) was added. The mixture was stirred at -78 °C for 15 min and then -23 °C for 120 h. After addition of saturated NaHCO₃, the resulting mixture was stirred at rt for 1 h and then filtered through a pad of Celite. The filtrate was extracted with dichloromethane. The combined organic layers were washed with brine, dried, and concentrated. The residue was passed through a short column of silica gel (n-hexane-ether = 6:1) to give a syrup, which was distilled under reduced pressure to give 21 (2.72 g, 71%) as a colorless liquid; bp 64 $^{\circ}$ C/19 mmHg (lit.^{31a} 80– 83 °C/7 mmHg for *ent*-21); $[\alpha]_{\rm D}^{21}$ +13.9 (*c* = 3.7, CCl₄) {lit.^{31b} $[\alpha]_{\rm D}^{25}$ +8.3 (*c* = 4.1, CCl₄), lit.^{31c} $[\alpha]_{\rm D}^{20}$ +12.3 (*c* = 2.0, CCl₄)}; IR (ZnSe) 3352, 3077, 2978, 2914, 1640, 1434, 1235, 1086, 992, 907 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 5.83 (2H, m), 5.14 (2H, m), 5.05 (1H, ddt, J = 17.1, 1.7, 1.6 Hz), 4.97 (1H, ddt, J = 10.3, 1.7, 1.3 Hz), 3.67 (1H, m), 2.29 (1H, m), 2.26-2.10 (3H, m), 1.61-1.52 (2H, m); ¹³C NMR (125 MHz, CDCl₃): δ 138.4, 134.7, 118.1, 114.8, 70.1, 41.9, 35.8, 30.0; HRMS (FI) calcd for C₈H₁₄O [M]⁺ 126.1045, found 126.1056.

(S)-Octa-1,7-dien-4-ol (*ent*-21). According to the procedure described above, allylation of 4-pentenal (2.56 g, 30.4 mmol) using (*R*)-(+)-l,l'-bi-2-naphthol (1.74 g, 6.08 mmol) instead of the S-isomer yielded *ent*-21 (2.86 g, 75%); $[\alpha]_{\rm D}^{21}$ –13.9 (*c* = 4.0, CCl₄) {lit.^{31a} $[\alpha]_{\rm D}^{25}$ –6.02 (*c* = 4.900, methanol)}; HRMS (FI) calcd for C₈H₁₄O [M]⁺ 126.1045, found 126.1059.

((2R,5R)-5-Allyltetrahydrofuran-2-yl)methanol (22). To a stirred suspension of 21 (909 mg, 7.20 mmol), Co(modp)₂ (776 mg, 1.44 mmol), and MS4A (0.92 g) in 2-propanol (67 mL) was added a 5.0-6.0 M solution of tert-butyl hydroperoxide in decane (1.50 mL), and the mixture was stirred at 50-52 °C for 6 h under an oxygen atmosphere, cooled to rt, and filtered through a pad of Celite. After addition of aqueous saturated Na2S2O3 to the filtrate, the resulting mixture was stirred at rt for 1 h, concentrated, diluted with dichloromethane, washed with brine, dried, and concentrated. The residue was chromatographed on silica gel (n-hexane-ethyl acetate = 2:1) to give 22 (765 mg, 75%) as a syrup; $[\alpha]_{\rm D}^{23}$ -7.3 (c 1.8, CHCl₃); IR (ZnSe) 3408, 3075, 2972, 2911, 1641, 1235, 1088, 882 cm^{-1} ; ¹ H NMR (500 MHz, CDCl₃): δ 5.79 (1H, ddt, J = 17.1, 10.1, 6.7 Hz), 5.07 (1H, ddt, J = 17.1, 2.0, 1.7 Hz), 5.04 (1H, ddt, J = 10.1, 2.0, 1.0 Hz), 4.10 (1H, ddd, J = 13.9, 6.4, 3.4 Hz), 4.01 (1H, ddt, J = 13.9, 6.1, 1.7 Hz), 3.61 (1H, dd, J = 11.5, 2.2 Hz), 3.47 (1H, dt, J = 11.5, 6.1 Hz), 2.38-2.32 (2H, m), 2.21 (1H, m), 2.03-1.92 (2H, m), 1.70–1.53 (2H, m); ¹³ C NMR (125 MHz, CDCl₃): δ 134.7, 117.0, 79.2, 78.6, 64.9, 40.0, 31.4, 27.4; HRMS (FI) calcd for C₈H₁₅O₂ [M + H] + 143.1072, found 143.1078.

((25,55)-5-Allyltetrahydrofuran-2-yl)methanol (*ent*-22). Treatment of *ent*-21 (918 mg, 7.27 mmol) as described above yielded *ent*-22 (733 mg, 71%); $[\alpha]_{\rm D}^{24}$ +7.1 (*c* 1.9, CHCl₃); HRMS (FI) calcd for C₈H₁₅O₂ [M + H]⁺ 143.1072, found 143.1064.

1-((2R,5R)-5-(Hydroxymethyl)tetrahydrofuran-2-yl)but-3-en-2-ol (7). To a stirred solution of **22** (470 mg, 3.31 mmol) in methanol (20 mL) was bubbled ozone (O₃) at -78 °C for 35 min. After the excess of O₃ was flushed out by the stream of nitrogen, dimethylsulfide (5.0 mL) was added. After stirring at -78 °C for 2 h and at -78 °C-rt for 4 h, the mixture was concentrated and coevaporated with benzene (×3) to give a syrup which was dissolved in tetrahydrofuran (10 mL). To the solution was added a 1.46 M solution of vinylmagnesium chloride (9.4 mL, 13.7 mmol) at 0 °C with stirring, and the mixture was stirred at 0 °C–rt for 2 h. After being quenched with saturated a queous NH₄Cl at 0 °C, the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried, and concentrated. The residue was chromatographed on silica gel (*n*-hexane–ethyl acetate = 2:1 \rightarrow 1:1 \rightarrow 0:1) to give 7 (470 mg, 83%) as a syrup; IR (ZnSe) 3340, 2932, 2872, 1647, 1419, 1318, 1218, 1037, 991, 916, 878 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.92–5.81 (1H, ms), 5.29–5.20 (1H, m), 5.14–5.03 (1H, m), 4.37–4.29 (1H, m), 4.25–4.09 (2H, m), 3.64–3.60 (1H, m), 3.50–3.45 (1H, m), 3.12 (2H, brs), 2.12–1.98 (2H, m), 1.77–1.54 (4H, m); ¹³ C NMR (125 MHz, CDCl₃): δ 140.8, 140.4, 114.3, 114.1, 79.6, 79.3, 79.2, 76.2, 72.5, 70.1, 64.7, 64.6, 42.4, 41.4, 32.7, 32.2, 27.1, 26.9; HRMS (ESI) calcd for C₉H₁₆O₃Na [M + Na] * 195.0997, found 195.0995.

1-((25,55)-5-(Hydroxymethyl)tetrahydrofuran-2-yl)but-3-en-2-ol (ent-7). According to the method described above, *ent-22* (237 mg, 1.67 mmol) was transformed into *ent-*7 (243 mg, 85%); HRMS (ESI) calcd for $C_9H_{16}O_3Na [M + Na]^+$ 195.0997, found 195.1001.

(5S)-3-(((2R,5R)-5-(2-((tert-Butvldimethylsilyl)oxy)but-3-en-1-yl)tetrahydrofuran-2-yl)methyl)-5-methyl-3-(phenylthio)dihydrofuran-2(3H)-one (24). To a stirred mixture of 7 (247 mg, 1.43 mmol) and 2,6-lutidine (1.73 mL, 14.8 mmol) in dichloromethane (10 mL) was added dropwise triflic anhydride (0.25 mL, 1.52 mmol) at -78 °C. After 1 h, tert-butyldimethylsilyl trifluoromethanesulfonate (0.43 mL, 1.86 mmol) was added and the reaction mixture was stirred at -78 °C for 1 h and then gradually warmed to 0 °C during 30 min. Crushed ice was added, and the resulting mixture was stirred for 20 min and extracted with ether. The combined organic layers were washed with cold aqueous HCl, water, saturated aqueous NaHCO3, water, and brine, dried, concentrated, and coevaporated with benzene $(\times 3)$ to give 23 (565 mg) which was employed for the next step without further purification. To a stirred solution of lithium hexamethyldisilazide prepared from a 1.65 M solution of nbutyllithium in n-hexane (2.6 mL, 4.29 mmol) and hexamethyldisilazane (0.90 mL, 4.29 mmol) in tetrahydrofuran (5.0 mL) was added dropwise a solution of 6 (894 mg, 4.29 mmol) in THF (3.0 mL) at -78 °C. After 7 min, the mixture was gradually warmed to 0 °C with stirring for 1 h. To this solution was added dropwise a solution of 23 (565 mg) in HMPA (2.0 mL) at -78 °C, and the resulting mixture was stirred at $-78 \rightarrow 0$ °C for 3 h. After addition of saturated aqueous NH4Cl, the resulting mixture was extracted with ether. The combined organic layers were washed with water and brine, dried, and concentrated. The residue was chromatographed on silica gel (*n*-hexane–ethyl acetate = $30:1 \rightarrow 20:1 \rightarrow 10:1 \rightarrow 4:1$) to give 24 (373 mg, 55% from 7) as a diastereomeric mixture.

IR (ZnSe) 3064, 2927, 2855, 1767, 1647, 1472, 1439, 1383, 1340, 1250, 1185, 1081, 833 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.57-7.52 (2H, m), 7.43-7.32 (3H, m), 5.86-5.76 (1H, m), 5.24-4.98 (2H, m), 4.66-4.50 (1H, m), 4.38 (0.82H, m), 4.22 (1H, m), 4.12-3.83 (1.18H, m), 2.99 (0.47H, dd, J = 14.2, 7.3 Hz), 2.90 (0.35H, dd, J = 14.2, 7.6 Hz), 2.88 (0.10H, dd, J = 14.1, 10.0 Hz), 2.81 (0.08H, dd, J = 14.0, 10.0 Hz), 2.26 (0.10H, dd, J = 14.1, 5.3 Hz), 2.10-1.42 (8.90H, m), 1.40 (0.30 H, d, J = 6.1 Hz), 1.39 (0.24H, d, J = 6.4 Hz), 1.21 (1.41H, d, J = 6.4 Hz), 1.15 (1.05H, d, J = 6.2 Hz), 0.89-0.88 (9H, m), 0.05–0.02 (12H, m); 13 C NMR (125 MHz, CDCl₃): δ 177.6, 177.4, 175.22, 175.15, 142.1, 141.0, 140.7, 137.3, 136.99, 136.96, 130.4, 130.3, 130.1, 129.71, 129.69, 129.3, 128.96, 128.94, 128.9, 114.5, 114.2, 113.3, 75.7, 75.6, 75.55, 75.32, 75.28, 74.4, 74.2, 73.8, 73.3, 73.1, 71.84, 71.79, 71.54, 71.52, 55.36, 55.33, 55.30, 55.27, 44.8, 44.4, 44.2, 41.9, 41.4, 41.2, 41.0, 40.9, 39.0, 38.8, 33.6, 33.5, 33.0, 32.7, 32.1, 31.9, 31.8, 29.7, 29.6, 29.3, 25.89, 25.87, 25.83, 21.3, 21.2, 20.73, 20.72, 18.23, 18.18, -4.26, -4.31, -4.41, -4.48, -4.83, -4.88, -4.93; HRMS (ESI) calcd for $C_{26}H_{40}O_4SSiNa [M + Na]^+$ 499.2314, found 499.2317.

(55)-3-(((25,55)-5-(2-((*tert*-Butyldimethylsilyl)oxy)but-3-en-1-yl)tetrahydrofuran-2-yl)methyl)-5-methyl-3-(phenylthio)-dihydrofuran-2(3*H*)-one (27). According to the procedure described above, *ent*-7 (244 mg, 1.42 mmol) was transformed into the corresponding lactone 27 (305 mg, 45% from *ent*-7).

IR (ZnSe) 3075, 2926, 2855, 1767, 1640, 1472, 1383, 1340, 1234, 1185, 1086, 865, 834 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.56–

The Journal of Organic Chemistry

7.48 (2H, m), 7.42–7.31 (3H, m), 5.86–5.75 (1H, m), 5.18–4.97 (2H, m), 4.63–3.85 (4H, m), 3.11–1.40 (10H, m), 1.39–1.11 (3H, m), 0.90–0.87 (9H, m), 0.06–0.015 (6H, m); ¹³C NMR (125 MHz, CDCl₃): δ 177.5, 177.4, 175.7, 141.9, 141.0, 140.9, 137.1, 137.0, 136.9, 130.48, 130,46, 129.9, 129.8, 129.4, 128.99, 128.96, 128.91, 114.22, 114.19, 113.5, 75.8, 75.7, 75.61, 75.57, 75.3, 75.1, 74.4, 74.0, 73.95, 73.87, 71.9, 71.6, 55.73, 55.65, 55.3, 44.8, 44.4, 44.3, 44.2, 43.04, 42.96, 41.9, 39.9, 39.7, 39.6, 33.73, 33.65, 32.7, 32.5, 32.1, 31.93, 31.87, 25.88, 25.87, 25.8, 21.49, 21.45, 20.7, 18.22, 18.19, –4.2, –4.4, –4.5, –4.8, –4.90, –4.93; HRMS (ESI) calcd for C₂₆H₄₀O₄SSiNa [M + Na]⁺ 499.2314, found 499.2321.

(3S,5S)-5-Methyl-3-(((2R,5R)-5-(2-oxobut-3-en-1-yl)tetrahydrofuran-2-yl)methyl)-3-(phenylthio)dihydrofuran-2(3H)-one (4a) and (3R,5S)-5-Methyl-3-(((2R,5R)-5-(2-oxobut-3en-1-yl)tetrahydrofuran-2-yl)methyl)-3-(phenylthio)dihydrofuran-2(3H)-one (4b). To a stirred solution of 24 (200 mg, 0.42 mmol) in dichloromethane (4.4 mL) was added a 10% HCl solution in methanol (2.2 mL) at rt. The mixture was stirred at rt for 1.2 h, made neutral by addition of NaHCO3 (powder), filtered through a pad of Celite, and then concentrated. The residue was diluted with ethyl acetate, washed with saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was passed through a short column of silica gel (*n*-hexane–ethyl acetate = $10:1 \rightarrow 4:1$) to give a syrup (143 mg) which was dissolved in dichloromethane-DMSO (1:1; 4.8 mL). Triethylamine (0.55 mL, 3.95 mmol) and a sulfur trioxide trimethylamine complex (274 mg, 1.97 mmol) were sequentially added to the solution at 0 °C, and the mixture was stirred at 0 °C-rt for 39 h, diluted with ether, and then washed with saturated aqueous NH₄Cl, water, and brine, dried, and concentrated. The residue was purified by preparative TLC (n-hexane-ethyl acetate = 4:1, 4 developments) to give 4a (108 mg, 72%) and 4b (13 mg, 8%).

4a. Syrup; $[\alpha]_{D}^{25}$ -80.6 (*c* 1.16, CHCl₃); IR (ZnSe) 3059, 2971, 2930, 1759, 1676, 1615, 1439, 1383, 1343, 1185, 1079, 974, 884, 753 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.55 (2H, brd, *J* = 7.4 Hz), 7.39 (1H, brt, *J* = 7.4 Hz), 7.33 (2H, brt, *J* = 7.4 Hz), 6.35 (1H, dd, *J* = 17.6, 10.5 Hz), 6.21 (1H, dd, *J* = 17.6, 1.0 Hz), 5.84 (1H, dd, *J* = 10.5, 1.0 Hz), 4.51 (1H, m), 4.44 (1H, m), 4.31 (1H, m), 2.88 (1H, dd, *J* = 14.2, 7.6 Hz), 2.87 (1H, dd, *J* = 15.4, 6.9 Hz), 2.66 (1H, dd, *J* = 15.4, 6.4 Hz), 2.11 (2H, m), 2.00 (1H, dd, *J* = 14.7, 9.8 Hz), 1.88 (1H, dd, *J* = 14.2, 6.8 Hz), 1.87 (1H, dd, *J* = 14.7, 2.7 Hz), 1.54 (2H, m), 1.18 (3H, *d*, *J* = 6.3 Hz); ¹³C NMR (125 MHz, CDCl₃): δ 198.8, 177.4, 136.9, 136.6, 130.2, 129.7, 128.9, 128.7, 74.7, 73.3, 55.0, 45.6, 41.2, 38.8, 32.3, 31.5, 21.2; HRMS (ESI) calcd for C₂₀H₂₄O₄SNa [M + Na]⁺ 383.1293, found 383.1288.

4b. Syrup; $[\alpha]_D^{25}$ +22.3 (*c* 0.22, CHCl₃); IR (ZnSe) 2972, 2919, 1759, 1676, 1384, 1188, 1086, 875, 753 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.52 (2H, brd, *J* = 7.4 Hz), 7.42 (1H, brt, *J* = 7.4 Hz), 7.34 (2H, brt, *J* = 7.4 Hz), 6.35 (1H, dd, *J* = 17.6, 10.5 Hz), 6.21 (1H, dd, *J* = 17.6, 1.0 Hz), 5.85 (1H, dd, *J* = 10.5, 1.0 Hz), 4.60 (1H, m), 4.41 (1H, m), 3.93 (1H, m), 2.88 (1H, dd, *J* = 15.7, 7.3 Hz), 2.74 (1H, dd, *J* = 14.1, 10.2 Hz), 2.60 (1H, dd, *J* = 15.7, 5.4 Hz), 2.23 (1H, dd, *J* = 14.1, 5.6 Hz), 2.13–1.95 (4H, m), 1.58–1.41 (2H, m), 1.34 (3H, d, *J* = 6.1 Hz); ¹³C NMR (125 MHz, CDCl₃): δ 198.9, 175.2, 137.3, 136.9, 130.1, 129.3, 128.9, 128.8, 76.2, 75.1, 74.0, 55.1, 45.5, 41.4, 40.5, 33.3, 31.7, 20.6; HRMS (ESI) calcd for C₂₀H₂₄O₄SNa [M + Na]⁺ 383.1293, found 383.1289.

(5*S*)-5-Methyl-3-(((2*S*,5*S*)-5-(2-oxobut-3-en-1-yl)tetrahydrofuran-2-yl)methyl)-3-(phenylthio)dihydrofuran-2(3*H*)-one (28). According to the procedure described above, 27 (174 mg, 0.37 mmol) was transformed into the corresponding allyl alcohol (121 mg, 91%). This compound (72 mg, 0.20 mmol) was oxidized as described above to give 28 (57.1 mg, 80%) as a light yellow oil; IR (ZnSe) 3057, 2973, 2919, 1758, 1676, 1615, 1439, 1383, 1342, 1185, 1085, 967, 878, 754 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.55–7.53 (2H, m), 7.49–7.30 (3H, m), 6.38 (0.4H, dd, *J* = 17.6, 10.5 Hz), 6.33 (0.6H, dd, *J* = 17.6, 10.0 Hz), 5.85 (0.4H, dd, *J* = 10.5, 1.0 Hz), 5.84 (0.6H, dd, *J* = 10.5, 1.0 Hz), 4.65 (0.4H, m), 4.58 (0.4H, m), 4.53 (0.6H, m), 4.41 (0.6H, m), 4.33 (0.4H, m), 3.96 (0.6H, m), 3.01 (0.6H, dd, *J* = 15.7, 7.6 Hz), 2.67 (0.4H, dd, J = 15.4, 5.9 Hz), 2.61 (0.6H, dd, J = 15.7, 5.1 Hz), 2.37 (0.4H, dd, J = 14.4, 5.3 Hz), 2.23 (0.4H, dd, J = 14.4, 10.2 Hz), 2.17–2.05 (2H, m), 2.04–1.93 (1.6H, m), 1.86 (0.6H, dd, J = 14.0, 6.6 Hz), 1.72 (0.4H, dd, J = 14.9, 10.0 Hz), 1.59–1.44 (2H, m), 1.37 (1.2H, d, J = 6.1 Hz), 1.08 (1.8H, d, J = 6.7 Hz); ¹³C NMR (125 MHz, CDCl₃): δ 198.9, 198.8, 177.4, 175.6, 137.1, 137.0, 136.8, 136.7, 130.4, 129.9, 129.8, 129.3, 129.0, 128.9, 128.8, 128.6, 76.1, 75.1, 74.7, 74.4, 74.1, 73.9, 55.5, 55.3, 45.7, 45.3, 42.8, 42.6, 39.8, 33.3, 32.2, 31.8, 31.7, 21.4, 20.6; HRMS (ESI) calcd for C₂₀H₂₄O₄SNa [M + Na]⁺ 383.1293, found 383.1288.

(3R,5S)-3-(((2R,5R)-5-((R,E)-8-((tert-Butyldimethylsilyl)oxy)-8-((2R,5R)-5-((R)-1-((tert-buty)dimethylsilyl)oxy)tridecyl)tetrahydrofuran-2-yl)-2-oxooct-3-en-1-yl)tetrahydrofuran-2yl)methyl)-5-methyl-3-(phenylthio)dihydrofuran-2(3H)-one (25). To a stirred mixture of lactone 4a (24.7 mg, 68.5 μ mol) and 3 (52.1 mg, 87.2 μ mol) in dichloromethane (2.0 mL) was added Grubbs' second-generation catalyst (10 mg, 11.8 μ mol). The mixture was stirred at rt for 5 min and at 40 °C for 2 h and then cooled to rt. Florisil was added with stirring, and the resulting mixture was filtered through a pad of Celite. The filtrate was concentrated to give a syrup, which was purified by preparative TLC (n-hexane-ethyl acetate = 2:1, 4 developments) to give 25 (43.1 mg, 68%) as a colorless oil; $[\alpha]_{\rm D}^{26}$ -17.7 (c 0.64, CHCl₃); IR (ZnSe) 2925, 2853, 1764, 1670, 1625, 1460, 1250, 1184, 1073, 832, 773 cm⁻¹; ¹ H NMR (500 MHz, $CDCl_3$): δ 7.54 (2H, brd, J = 7.1 Hz), 7.39 (1H, brt, J = 7.1 Hz), 7.34 (2H, brt, J = 7.1 Hz), 6.82 (1H, dt, J = 15.9, 6.8 Hz), 6.10 (1H, dt, J = 15.9, 1.5 Hz), 4.52 (1H, m), 4.46 (1H, m), 4.30 (1H, m), 3.90 (2H, m), 3.60 (1H, m), 3.56 (1H, m), 2.91 (1H, dd, J = 14.2, 7.6 Hz), 2.85 (1H, dd, J = 15.4, 6.4 Hz), 2.61 (1H, dd, J = 15.4, 6.6 Hz), 2.22 (2H, m), 2.11 (2H, m), 1.99 (1H, brdd, J = 14.7, 9.8 Hz), 1.88 (1H, dd, J = 14.2, 6.3 Hz), 1.87 (1H, brdd, J = 14.7, 2.9 Hz), 1.83 (2H, m), 1.68-1.22 (30H, m), 1.20 (1H, dd, J = 6.3 Hz), 0.87 (3H, t, J = 6.8 Hz), 0.87 (18H, s), 0.06 (3H, s), 0.05 (3H, s), 0.04 (6H, s); ¹³ C NMR (125 MHz, CDCl₃): δ 198.4, 177.4, 147.9, 137.0, 130.5, 130.3, 129.7, 128.9, 81.8, 81.7, 74.9, 74.73, 74.65, 74.4, 73.3, 55.0, 46.0, 41.1, 39.0, 32.71, 32.68, 32.4, 32.2, 31.9, 31.6, 29.8, 29.7, 29.63, 29.62, 29.61, 29.6, 29.3, 27.3, 25.94, 25.93, 25.8, 24.4, 22.7, 21.3, 18.18, 18.17, 14.1, -4.28, -4.30, -4.5, -4.6; HRMS (ESI) calcd for C₅₃H₉₂O₇SSi₂Na [M + Na] ⁺ 951.6000, found 951.6006.

(5S)-3-(((2S,5S)-5-((R,E)-8-((tert-Butyldimethylsilyl)oxy)-8-((2R,5R)-5-((R)-1-((tert-butyldimethylsilyl)oxy)tridecyl)tetrahydrofuran-2-yl)-2-oxooct-3-en-1-yl)tetrahydrofuran-2-yl)methyl)-5-methyl-3-(phenylthio)dihydrofuran-2(3H)-one (29). Treatment of lactone 28 (25.0 mg, 69.4 µmol) and 3 (53.1 mg, 88.9 μ mol) with Grubbs' second-generation catalyst (10.4 mg, 12.3 μ mol) as described for preparation of 25 gave 29 (47.1 mg, 73%) as a colorless oil; IR (ZnSe) 2924, 2853, 1763, 1670, 1625, 1463, 1439, 1386, 1251, 1185, 1073, 833, 773 cm⁻¹; ¹H NMR (500 MHz, CDCl₂): δ 7.55–7.53 (1.30H, brd, J = 7.1 Hz), 7.49–7.47 (0.70H, brd, J = 7.1Hz), 7.41-7.31 (3H, m), 6.83 (0.35H, dt, J = 15.9, 6.9 Hz), 6.80 (0.65H, dt, J = 15.9, 6.8 Hz), 6.11 (0.35H, brd, J = 15.9, 1.5 Hz), 6.07 (0.65H, brd, J = 15.9, 1.5 Hz), 4.64 (0.35H, m), 4.58 (0.35H, m), 4.54 (0.65H, m), 4.40 (0.65H, m), 4.31 (0.35H, m), 3.95 (0.65H, m), 3.89 (2H, m), 3.59 (1H, m), 3.57 (1H, m), 3.02 (0.65H, dd, J = 14.0, 7.9 Hz), 2.88 (0.35H, dd, J = 15.4, 6.8 Hz), 2.82 (0.65H, dd, J = 15.7, 7.4 Hz), 2.62 (0.35H, dd, J = 15.4, 6.3 Hz), 2.57 (0.65H, dd, J = 15.7, 5.6 Hz), 2.38 (0.35H, dd, J = 14.4, 5.4 Hz), 2.23-1.22 (39H, m), 1.37 (1.05H, d, J = 6.2 Hz), 1.07 (1.95H, d, J = 6.4 Hz), 0.89-0.87 (21H, J)m), 0.06, 0.053, 0.045, 0.041, 0.034, 0.028 (total 12H, each s); ¹³C NMR (125 MHz, CDCl₃): δ 198.5, 198.4, 177.4, 175.6, 148.0, 147.9, 137.1, 137.0, 130.6, 130.5, 129.9, 129.8, 129.4, 129.0, 128.9, 81.8, 81.7, 76.0, 75.2, 74.9, 74.7, 74.4, 74.3, 74.1, 73.9, 55.5, 55.4, 46.1, 45.8, 43.0, 42.7, 39.83, 39.77, 33.3, 32.72, 32.69, 32.3, 32.1, 31.9, 31.8, 31.7, 29.8, 29.7, 29.63, 29.62, 29.61, 29.59, 29.33, 27.3, 27.2, 25.94, 25.91, 25.8, 24.5, 24.4, 22.7, 21.4, 20.6, 18.20, 18.17, 14.1, -4.3, -4.5, -4.6; HRMS (ESI) calcd for $C_{53}H_{92}O_7SSi_2Na$ [M + Na]⁺ 951.6000, found 951.6025.

(3R,5S)-3-(((2R,5R)-5-((R)-8-((tert-Butyldimethylsilyl)oxy)-8-((2R,5R)-5-((R)-1-((tert-butyldimethylsilyl)oxy)tridecyl)-tetrahydrofuran-2-yl)-2-oxooctyl)tetrahydrofuran-2-yl)-methyl)-5-methyl-3-(phenylthio)dihydrofuran-2(3H)-one (26).

The Journal of Organic Chemistry

A mixture of 25 (20.4 mg, 21.9 μ mol) and PtO₂ (5.6 mg) in THF (1.1 mL) was vigorously stirred at rt under a hydrogen atmosphere for 13 h, filtered through a pad of Celite, and concentrated. The residue was purified by preparative TLC (*n*-hexane-ethyl acetate = 10:1, 6developments) to give 26 (17.1 mg, 84%) as a colorless oil; $\left[\alpha\right]_{D}^{24} =$ -17.7 (c 0.66, CHCl₃); IR (ZnSe) 2924, 2854, 1767, 1713, 1462, 1250, 1185, 1082, 833, 773 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 7.56-7.54 (2H, m), 7.41-7.32 (1H, m), 7.37-7.32 (2H, m), 4.51 (1H, m), 4.44 (1H, m), 4.29 (1H, m), 3.90 (2H, m), 3.56 (2H, m), 2.88 (1H, dd, J = 14.2, 7.6 Hz), 2.62 (1H, dd, J = 15.2, 7.6 Hz), 2.48 (1H, dd, J = 15.2, 5.6 Hz), 2.43 (2H, dt, J = 7.4, 1.2 Hz), 2.11 (2H, m), 1.99 (1H, dd, J = 14.7, 10.0 Hz), 1.88 (1H, dd, J = 14.2, 6.8 Hz), 1.87 (1H, dd, J = 14.7, 2.2 Hz), 1.82 (2H, m), 1.67–1.25 (34H, m), 1.18 (3H, d, J = 6.3 Hz), 0.876 (3H, t, J = 6.9 Hz), 0.875 (9H, s), 0.870(9H, s), 0.049 (3H, s), 0.046 (3H, s), 0.042 (3H, s), 0.03 (3H, s); ¹³ C NMR (125 MHz, CDCl₃): δ 209.4, 177.4, 137.0, 130.3, 129.8, 129.0, 81.8, 81.7, 74.8, 74.74, 74.71, 73.3, 55.0, 48.9, 43.1, 41.3, 38.9, 32.6, 32.4, 32.3, 31.9, 31.6, 29.8, 29.7, 29.65, 29.63, 29.62, 29.61, 29.5, 29.3, 27.3, 27.2, 25.95, 25.87, 25.8, 23.6, 22.7, 21.3, 18.2, 14.1, -4.3, -4.6; HRMS (ESI) calcd for $C_{53}H_{94}O_7SSi_2Na [M + Na]^+ 953.6156$, found 953.6166.

(5S)-3-(((2S,5S)-5-((R)-8-((tert-Butyldimethylsilyl)oxy)-8-((2R,5R)-5-((R)-1-((tert-butyldimethylsilyl)oxy)tridecyl)tetrahydrofuran-2-yl)-2-oxooctyl)tetrahydrofuran-2-yl)methyl)-5methyl-3-(phenylthio)dihydrofuran-2(3H)-one (30). Treatment of 29 (24.3 mg, 26.1 μ mol) with PtO₂ (6.6 mg) as described for preparation of 26 gave 30 (20.3 mg, 84%) as a colorless oil; IR (ZnSe) 2925, 2854, 1764, 1715, 1463, 1383, 1251, 1185, 1084, 832, 773 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.56–7.54 (1.30H, m), 7.49–7.47 (0.70H, m), 7.42-7.31 (3H, m), 4.64 (0.35H, m), 4.56 (0.35H, m), 4.53 (0.65H, m), 4.36 (0.65H, m), 4.28 (0.35H, m), 3.96 (0.65H, m), 3.89 (2H, m), 3.58 (2H, m), 3.01 (0.65H, dd, J = 14.0, 7.8 Hz), 2.66 (0.35H, dd, J = 15.4, 7.6 Hz), 2.64 (0.65H, dd, J = 15.9, 7.8 Hz), 2.48 (0.35H, dd, J = 15.4, 5.4 Hz), 2.46 (0.65H, dd, J = 15.9, 5.2 Hz), 2.45 (0.70H, t, J = 6.9 Hz), 2.39 (1.30H, t, J = 7.1 Hz), 2.36 (0.35H, dd, J = 14.2, 5.4 Hz), 2.23 (0.35H, dd, J = 14.2, 10.0 Hz), 2.11-1.22 (40.65H, m), 1.37 (1.05H, d, J = 6.1 Hz), 1.09 (1.95H, d, J = 6.4 Hz), 0.89–0.86 (21H, m), 0.052, 0.048, 0.044, 0.041, 0.038, 0.030 (total 12H, each s); ¹³C NMR (125 MHz, CDCl₃): δ 209.4, 209.2, 177.3, 175.6, 137.1, 137.0, 130.5, 129.9, 129.4, 129.0, 128.9, 81.8, 81.7, 76.1, 76.0, 74.9, 74.8, 74.7, 74.6, 74.5, 74.4, 74.1, 74.0, 73.90, 73.87, 55.5, 55.3, 48.9, 48.6, 43.6, 43.4, 42.8, 42.7, 39.9, 33.3, 32.6, 32.4, 32.3, 31.9, 31.7, 29.8, 29.6, 29.5, 29.3, 27.3, 25.96, 25.94, 23.6, 22.7, 21.5, 21.4, 18.2, 14.1, -4.27, -4.31, -4.5, -4.6; HRMS (ESI) calcd for C₅₃H₉₄O₇SSi₂Na [M + Na]⁺ 953.6156, found 953.6149.

(S)-3-(((2R,5R)-5-((R)-8-Hydroxy-8-((2R,5R)-5-((R)-1-hydroxytridecyl)tetrahydrofuran-2-yl)-2-oxooctyl)tetrahydrofuran-2yl)methyl)-5-methylfuran-2(5H)-one (1). To a stirred solution of 26 (16.3 mg, 17.4 μ mol) in dichloromethane (0.9 mL) was added mCPBA (70-75% assay; 4.3 mg) at 0 °C. After 30 min, aqueous saturated NaHCO₃/Na₂S₂O₃ (1:1) was added, and the resulting mixture was extracted with ether. The combined organic layers were washed with aqueous saturated NaHCO₃, water, and brine, dried, and concentrated. The residue (22.4 mg) was dissolved in toluene (0.9 mL). The solution was heated at 100-105 °C for 1.5 h with stirring and concentrated to give a crude butenolide (18.5 mg). To a stirred solution of the butenolide in dichloromethane (0.8 mL) was added HF·pyridine (80 μ L) at 0 °C, and the mixture was stirred at 0 °C for 20 min. After addition of aqueous saturated NaHCO₃, the resulting mixture was stirred at 0 °C for 10 min and extracted with ethyl acetate. The combined organic layers were washed with aqueous saturated NaHCO₃, water, and brine, dried, and concentrated. The residue was purified by preparative TLC (ethyl acetate, 3 developments) to give 1 (9.0 mg, 87% from 26) as a white powder; mp 72-73 °C (n-hexaneether); $[\alpha]_D^{25} = +1.4$ (c 0.42, CHCl₃); IR (ZnSe) 3447, 2917, 2849, 1751, 1734, 1700, 1456, 1404, 1373, 1323, 1270, 1202, 1027 cm⁻¹; HRMS (ESI) calcd for $C_{35}H_{60}O_7Na$ [M + Na]⁺ 615.4237, found 615.4239.

(S)-3-(((25,5S)-5-((R)-8-Hydroxy-8-((2R,5R)-5-((R)-1-hydroxytridecyl)tetrahydrofuran-2-yl)-2-oxooctyl)tetrahydrofuran-2**yl)methyl)-5-methylfuran-2(5***H***)-one (31).** According to the procedure described for preparation of 1 from 26, compound 30 (18.8 mg, 20.1 μ mol) was transformed into 31 (10.4 mg, 87%) as a white powder; mp 58–60 °C (*n*-hexane-ether); $[\alpha]_D^{25} = +41.2$ (*c* 0.61, CHCl₃); IR (ZnSe) 3465, 2915, 2849, 1751, 1734, 1700, 1467, 1413, 1342, 1317, 1197, 1024, 961 cm⁻¹; HRMS (ESI) calcd for C₃₅H₆₀O₇Na [M + Na]⁺ 615.4237, found 615.4236.

Montanacin D-15,20-Diacetate (33). According to the procedure described in the literature,⁵ 32 (1.0 mg, 1.7 μ mol) was transformed into 33 (0.9 mg, 79%) as a syrup; $[\alpha]_{\rm D}^{27}$ +17.5 (*c* 0.09, CHCl₃); IR (ZnSe) 2931, 2855, 2922, 1749, 1455, 1369, 1200, 1027 cm⁻¹; HRMS (ESI) calcd for C₃₉H₆₄O₉Na [M + Na] ⁺ 699.4448, found 699.4447.

Bis-TMS Derivative 34 of 32. According to the procedure described in the literature, ⁵ 32 (0.9 mg, 1.5 μ mol) was transformed into 34 (0.9 mg, 81%) as an amorphous solid; ¹H NMR (500 MHz, CDCl₃): δ 6.66 (1H, m), 4.45 (1H, brq, J = 6.8 Hz), 4.01–3.94 (2H, m), 3.66 (1H, m), 3.62 (1H, m), 3.60 (1H, m), 3.33 (1H, m), 2.37 (1H, dd, J = 15.6, 8.8 Hz), 2.33–2.23 (2H, m), 2.17–2.13 (2H, m), 1.94 (1H, dd, J = 15.6, 3.9 Hz), 1.75–1.17 (38H, m), 0.99 (3H, d, J = 6.8 Hz), 0.91 (3H, t, J = 6.6 Hz), 0.25, (18H, s); HRMS (EI) calcd for C₄₁H₇₆O₇Si₂ [M]⁺ 736.5130, found 736.5131.

Bis-TMS Derivative 35 of 1. According to the procedure described in the literature, ⁵ 1 (1.0 mg, 1.7 μ mol) was transformed into **35** (1.2 mg, 96%) as an amorphous solid; ¹H NMR (500 MHz, CDCl₃): δ 6.50 (1H, m), 4.29–4.26 (2H, m), 4.02 (1H, m), 3.98 (2H, m), 3.64 (1H, m), 3.61 (1H, m), 2.41 (1H, dd, J = 15.4, 7.3 Hz), 2.34–2.26 (2H, m), 2.13 (2H, t, J = 7.3 Hz), 2.07 (1H, dd, J = 15.4, 5.6 Hz), 1.78–1.12 (38H, m), 0.91 (3H, t, J = 6.9 Hz), 0.86 (3H, d, J = 6.9 Hz), 0.244 (9H, s), 0.242, (9H, s); HRMS (EI) calcd for C₄₁H₇₆O₇Si₂ [M]⁺ 736.5130, found 736.5110.

(3S)-3-((tert-Butyldimethylsilyl)oxy)nona-1,8-dien-5-ol (36). To a stirred solution of (S)-3-TBSoxy-4-pentenal (0.79 g, 3.68 mmol) in tetrahydrofuran (20 mL) was added dropwise a 0.5 M solution of 3-butenylmagnesium bromide in ether (10 mL, 5.0 mmol) at -78 °C, and the mixture was stirred at -78 °C for 1.5 h and at 0 °C for 1 h. Saturated aqueous NH₄Cl was added with vigorously stirring, and then the resulting mixture was extracted with ether. The combined organic layers were washed successively with water and brine, dried, and concentrated. The residue was chromatographed on silica gel (nhexane-ether = $30:1 \rightarrow 10:1$) to give 36 (884 mg, 89%) as a syrup; IR (ZnSe) 3441, 3079, 2928, 2857, 1641, 1471, 1254, 1084, 910, 835, 775 cm⁻¹; ¹ H NMR (500 MHz, CDCl₃): δ 5.90–5.79 (2H, m), 5.27–4.94 (4H, m), 4.52 (0.44H, m), 4.37 (0.56H, m), 3.92 (0.44H, m), 3.84 (0.56H, m), 3.38 (0.44H, brs), 3.21 (0.56H, brs), 2.22-2.09 (2H, m), 1.75-1.43(4H, m), 0.912 (3.96H, s), 0.906 (5.04H, s), 0.11 (1.32H, s), 010 (1.68H, s), 0.07 (3H, s); 13 C NMR (125 MHz, CDCl₃): δ 141.4, 140.0, 138.59, 138.57, 114.5, 114.4, 75.0, 72.7, 70.2, 67.9, 44.4, 43.0, 36.8, 36.7, 29.8, 29.6, 25.80, 25.79, 18.1, 18.0, -3.9, -4.6, -4.9, -5.2; HRMS (FI) calcd for $C_{15}H_{31}O_2Si [M + H]^+ 271.2093$, found 271.2090.

(4R,6S)-4-(But-3-en-1-yl)-2-phenyl-6-vinyl-1,3-dioxane (37) and (45,65)-4-(But-3-en-1-yl)-2-phenyl-6-vinyl-1,3-dioxane (38). To a stirred solution of 36 (1.43 g, 5.28 mmol) in tetrahydrofuran (6.0 mL) was added a 1.0 M solution of tetrabutylammonium fluoride in tetrahydrofuran (6.34 mL) at 0 °C, and the mixture was stirred at 0 $^\circ C \rightarrow$ rt for 3 h, diluted with ethyl acetate, washed with brine, dried, and concentrated. The residue was passed through a short column of silica gel (*n*-hexane–ethyl acetate = $3:1 \rightarrow 0:1$) to give a syrup (744 mg), which was dissolved in N,Ndimethylformamide (5.0 mL). To the solution were added benzaldehyde dimethylacetal (0.86 mL, 5.71 mmol) and dcamphorsulfonic acid (55 mg, 0.48 mmol). The mixture was stirred under diminished pressure (~2.6 kPa) at 50 °C for 1 h, cooled, diluted with ether, washed successively with saturated aqueous NaHCO₃, water, and brine, dried, and concentrated. The residue was chromatographed on silica gel (*n*-hexane-dichloromethane = $6:1 \rightarrow$ $4:1 \rightarrow 2:1 \rightarrow n$ -hexane-ethyl acetate = 4:1 and then *n*-hexanedichloromethane = $8:1 \rightarrow 7:1 \rightarrow 6:1$) to give 37 (483 mg, 37% from 36) and 38 (511 mg, 40% from 36).

The Journal of Organic Chemistry

37 (major/minor = ca. 2.3:1). Syrup; IR (ZnSe) 3072, 3032, 2977, 2922, 1639, 1399, 1235, 1088, 991, 903, 743 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.53–7.50 (2H, m), 7.38–7.26 (3H, m), 6.11 (0.73H, ddd, J = 17.7, 11.0, 4.0 Hz), 5.96–5.80 (1.27H, m), 5.83 (0.27H, s), 5.82 (0.73H, s), 5.40 (0.73H, dt, J = 11.0, 1.2 Hz), 5.35 (0.27H, dt, J = 17.3, 1.0 Hz), 5.34 (0.73H, dt, J = 17.7, 1.7 Hz), 5.18 (0.27H, dt, J = 10.6, 1.2 Hz), 5.08 (0.27H, dt, J = 17.7, 1.5 Hz), 5.04 (0.73H, dt, J = 10.3, 1.5 Hz), 4.98 (0.73H, dt, J = 10.3, 1.4 Hz), 4.82 (0.73H, brs), 4.55 (0.27H, m), 4.27 (0.27H, m), 3.97 (0.73H, m), 2.34–2.03 (3H, m), 1.81–1.54 (3H, m); ¹³C NMR (125 MHz, CDCl₃): δ 139.0, 138.9, 138.2, 138.1, 137.9, 137.3, 128.7, 128.6, 128.2, 126.1, 126.0, 117.6, 115.6, 115.1, 114.9, 95.1, 93.9, 73.0, 72.7, 72.05, 71.99, 35.1, 33.8, 33.3, 30.0, 29.7, 29.2; HRMS (EI) calcd for C₁₆H₂₀O₂ [M]⁺ 244.1463, found 244.1442.

38. Syrup; $[\alpha]_D^{24} - 15.7$ (*c* 0.96, CHCl₃); IR (ZnSe) 3070, 3032, 2977, 2915, 2844, 1640, 1336, 1149, 1088, 1017, 902, 840, 756 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.56 (2H, brd, *J* = 6.8 Hz), 7.40–7.32 (3H, m), 5.97 (1H, ddd, *J* = 17.3, 10.3, 5.4 Hz), 5.87 (1H, ddt, *J* = 17.1, 10.3, 6.6 Hz), 5.60 (1H, s), 5.37 (1H, dt, *J* = 17.3, 1.4 Hz), 5.20 (1H, dt, *J* = 10.5, 1.2 Hz), 5.08 (1H, dt, *J* = 17.1, 1.5 Hz), 5.02 (1H, dt, *J* = 10.3, 0.8 Hz), 4.37 (1H, m), 3.89 (1H, m), 2.32–2.19 (2H, m), 1.81 (1H, ddd, *J* = 13.7, 8.8, 5.8 Hz), 1.69 (1H, dt, *J* = 13.2, 2.4 Hz), 1.66 (1H, m), 1.56 (1H, dt, *J* = 13.2, 11.3 Hz); ¹³C NMR (125 MHz, CDCl₃): δ 138.7, 138.1, 137.8, 128.6, 128.1, 126.1, 115.4, 114.8, 100.5, 77.2, 75.9, 36.6, 34.9, 29.1; HRMS (EI) calcd for C₁₆H₂₀O₂ [M]⁺ 244.1463, found 244.1476.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b02187.

NMR spectra of 1, 3, 4a, 4b, 5, 7-9, 11-14, 16, 17, 19-

22, 24-31, 33, and 36-38 (PDF)

2D-NMR spectra of 1, 31, and 33; MS spectra of 34 and 35, CAST/CNMR data (PDF)

AUTHOR INFORMATION

Corresponding Author

*E-mail: shunyat@riken.jp.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by JSPS KAKENHI Grant Numbers 24580168, 16K07724.

REFERENCES

(1) For recent reviews, see: (a) Qayed, W. S.; Aboraia, A. S.; Abdel-Rahman, H. M.; Youssef, A. F. *Pharm. Chemica* **2015**, *7*, 24–35. (b) Choo, C.-Y.; Abdullah, N.; Diederich, M. *Phytochem. Rev.* **2014**, *13*, 835–851. (c) Smith, R. E.; Tran, K.; Richard, K. M. In *Studies in Natural Product Chemistry*; Attra-ur-Rahman, Ed.; Elsevier B. V., Oxford, 2014; Vol. *41*, pp 95–117. (d) Liaw, C.-C.; Wu, T.-Y.; Chang, F.-R.; Wu, Y.-C. *Planta Med.* **2010**, *76*, 1390–1404. (e) Spurr, I. B.; Brown, R. C. D. *Molecules* **2010**, *15*, 460–501. (f) Alali, F. Q.; Liu, X.-X.; McLaughlin, J. L. J. Nat. Prod. **1999**, *62*, 504–540. (g) Zafra-Polo, M. C.; Figadere, B.; Gallardo, T.; Tormo, J. R.; Cortes, D. *Phytochemistry* **1998**, *48*, 1087–1117 and references cited therein.

(2) Wang, L.-Q.; Zhao, W.-M.; Qin, G.-W.; Cheng, K.-F.; Yang, R.-Z. Nat. Prod. Lett. 1999, 14, 83–90.

(3) Wang, L.-Q.; Nakamura, N.; Meselhy, M. R.; Hattori, M.; Zhao, W.-M.; Cheng, K.-F.; Yang, R.-Z.; Qin, G.-W. *Chem. Pharm. Bull.* **2000**, *48*, 1109–1113.

(4) Chen, C. -Y.; Chang, F.-R.; Chiu, H.-F.; Wu, M.-J.; Wu, Y.-C. *Phytochemistry* **1999**, *51*, 429–433.

(5) Alfonso, D.; Colman-Saizarbitoria, T.; Zhao, G.-X.; Shi, G.; Ye, Q.; Schwedler, J. T.; McLaughlin, J. L. *Tetrahedron* **1996**, *52*, 4215–4224.

(6) Gu, Z.-M.; Zhao, G.-X.; Oberlies, N. H.; Zeng, L.; McLaughlin, J. L. In *Recent Advances in Phytochemistry*; Romeo, J. T., Ed.; Plenum Press: New York, 1995, Vol. 29, pp 249–310.

(7) Shi, G.; Zeng, L.; Gu, Z.-M.; MacDougal, J. M.; McLaughlin, J. L. *Heterocycles* **1995**, *41*, 1785–1796.

(8) Fujimoto, Y.; Murasaki, C.; Shimada, H.; Nishioka, S.; Kakinuma, K.; Singh, S.; Singh, M.; Gupta, Y. K.; Sahai, M. *Chem. Pharm. Bull.* **1994**, *42*, 1175–1184.

(9) (a) Takahashi, S.; Hongo, Y.; Tsukagoshi, Y.; Koshino, H. Org. Lett. **2008**, *10*, 4223–4226. (b) Takahashi, S.; Takahashi, R.; Hongo, Y.; Koshino, H.; Yamaguchi, K.; Miyagi, T. J. Org. Chem. **2009**, *74*, 6382–6385 and references cited therein.

(10) White, J. D.; Somers, T. C.; Reddy, G. N. J. Org. Chem. 1992, 57, 4991-4998.

(11) Hartung, J.; Greb, M. J. Organomet. Chem. 2002, 661, 67-84.

(12) Inoki, S.; Mukaiyama, T. Chem. Lett. 1990, 19, 67-70.

(13) Palmer, C.; Morra, N. A.; Stevens, A. C.; Bajtos, B.; Machin, B. P.; Pagenkopf, B. L. Org. Lett. **2009**, *11*, 5614–5617.

(14) Two sequential Pd-catalyzed cyclization reactions of a monoprotected 1,2-diol derivative were reported: Ward, A. F.; Wolfe, J. P. Org. Lett. 2009, 11, 2209–2212.

(15) Several papers have appeared dealing with an oxidative cyclization of a monoprotected alcohol of diols: (a) Wang, J.; Pagenkopf, B. L. Org. Lett. 2007, 9, 3703–3706. (b) Li, Y.; Zhou, F.; Forsyth, C. J. Angew. Chem., Int. Ed. 2007, 46, 279–282. (c) Chen, Z.; Sinha, S. C. Tetrahedron Lett. 2009, 50, 6398–6401 and references cited therein.

(16) For construction of a bis-THF ring by double cyclization of a 1,2-diol derivative, see: Wang, Z.-M.; Tian, S.-K.; Shi, M. *Eur. J. Org. Chem.* **2000**, 2000, 349–356.

(17) Kotsuki, H.; Kadota, I.; Ochi, M. J. Org. Chem. **1990**, 55, 4417–4422.

(18) Mukai, C.; Sonobe, H.; Kim, J. S.; Hanaoka, M. J. Org. Chem. 2000, 65, 6654–6659.

(19) The corresponding *cis*-isomer could not be isolated.

(20) Fries, P.; Müller, M. K.; Hartung, J. Tetrahedron 2014, 70, 1336-1347.

(21) Both compounds were prepared from (S)-3-TBSoxy-4-pentenal (refs 22, 23) *via* the corresponding benzylidene derivatives **37** and **38** (see Experimental Section) (ref 24-27).

(22) Hatakeyama, S.; Okano, T.; Maeyama, J.; Esumi, T.; Hiyamizu, H.; Iwabuchi, Y.; Nakagawa, K.; Ozono, K.; Kawase, A.; Kubodera, N. *Bioorg. Med. Chem.* **2001**, *9*, 403–415.

(23) Sabitha, G.; Srinivas, R.; Yadav, J. S. Synthesis **2011**, 2011, 3343–3349.

(24) We initially prepared the corresponding acetonides for determination (ref 25) of the stereochemistry, but the separation failed.

(25) (a) Rychnovsky, S. D.; Skalitzky, D. J. Tetrahedron Lett. **1990**, 31, 945–948. (b) Evans, D. A.; Rieger, D. L.; Gage, J. R. Tetrahedron Lett. **1990**, 31, 7099–7100. (c) Rychnovsky, S. D.; Richardson, T. I.; Rogers, B. N. J. Org. Chem. **1997**, 62, 2925–2934.

(26) The ¹H NMR spectrum of the less polar isomer on TLC (*n*-hexane-dichloromethane = 2:1) showed it to be a single product while the more polar one was revealed to be composed of a 1:2.3 mixture of two stereoisomers by the ¹H NMR analysis; the stereochemistry was determined by NOESY spectra. Therefore, the former and latter were determined to be *cis*-benzylidene derivative **38** and *trans*-isomer **37**, respectively.

(27) For several new synthetic methods for preparation of a similar protected 1,3-syn diol derivative, see; (a) Bredenkamp, A.; Zhu, Z.-B.; Kirsch, S.-F. Eur. J. Org. Chem. 2016, 2016, 252-254. (b) Goodwin, J. A.; Ballesteros, C. F.; Aponick, A. Org. Lett. 2015, 17, 5574-5577. (c) Herrmann, A. T.; Saito, T.; Stivala, C. E.; Tom, J.; Zakarian, A. J. Am. Chem. Soc. 2010, 132, 5962-5963 and references cited therein.

(28) In the ¹H NMR spectra, the splitting pattern of the olefinic methine proton was clearly different between the major and minor compound.

(29) Attempted oxidative cyclization of 36 (syn/anti = ca. 1.2/1) under Pagenkopf's condition afforded a mixture of four compounds which were separated as the corresponding diacetates: 16 (14% yield from 36), 17 (15%), 19 (25%), and 20 (13%). Mukaiyama's cyclization also gave a similar result: 16 (13% yield from 36), 17 (15%), 19 (21%), and 20 (12%).

(30) Keck, G. E.; Tarbet, K. H.; Geraci, L. S. J. Am. Chem. Soc. 1993, 115, 8467-8468.

(31) (a) Bartlett, P. A.; Meadows, J. D.; Ottow, E. J. Am. Chem. Soc. 1984, 106, 5304–5311. (b) Bartlett, P. A.; Johnson, W. S.; Elliott, J. D. J. Am. Chem. Soc. 1983, 105, 2088–2089. (c) Johnson, W. S.; Crackett, P. H.; Elliott, J. D.; Jagodzinski, J. J.; Lindell, S. D.; Natarajan, S. Tetrahedron Lett. 1984, 25, 3951–3954.

(32) The enantiomeric purity of alcohol **21** or its enantiomer was determined by the 1 H NMR analysis (500 MHz) of the corresponding MTPA esters.

(33) For preparation of (\pm) -22 (*trans/cis* = 93/3) *via* addition of allyltributyltin to 2,7-dioxabicyclo[2.2.1]heptane, see: Friestad, G. K.; Lee, H. J. Org. Lett. 2009, 11, 3958–3961. Synthesis of the corresponding (–)-*cis*-isomer was also disclosed: Pilli, R. A.; Riatto, V. B. Tetrahedron: Asymmetry 2000, 11, 3675–3686.

(34) Different from the case of the THP analog, the use of the sodium enolate in this coupling reaction gave unsatisfactory results. The use of *N*-methylpyrrolidone instead of HMPA gave a complex mixture.

(35) The stereochemistry of each isomer was deduced by the ¹H NMR analyses. In the ¹H NMR spectra, the methyl group of **4a** was observed at δ 1.18 (d, J = 6.3 Hz) while the minor isomer **4b** showed the corresponding signal as a doublet (J = 6.1 Hz) at 1.34 ppm. The data are consistent with those reported in the literature: Sinha, S. C.; Keinan, E. J. Am. Chem. Soc. **1993**, 115, 4891–4892. Sinha, A.; Sinha, S. C.; Sinha, S. C.; Keinan, E. J. Org. Chem. **1999**, 64, 2381–2386 and references cited therein.

(36) Grubbs, R. H. Handbook of Metathesis, Vol. 1-3; Wiley-VCH: 2003.

(37) This lactone was an inseparable mixture regarding the SPh group.

(38) Gu, Z.-M.; Zhao, G. X.; Oberlies, N. H.; Zeng, L.; McLaughlin, J. L. In *Phytochemistry of Medicinal Plants*; Arnason, J. T., Mata, R.,

Romeo, J. T., Eds.; Plenum Press: New York, 1995; pp 249–310. (39) Satoh, H.; Koshino, H.; Uzawa, J.; Nakata, T. *Tetrahedron* 2003,

(c)) Gateri, 1., 100 mill, 11, 02 and 1, 1, 10 and 1, 120

(40) Satoh, H.; Koshino, H.; Uno, T.; Koichi, S.; Iwata, S.; Nakata, T. *Tetrahedron* **2005**, *61*, 7431–7437.

(41) Koichi, S.; Iwata, S.; Uno, T.; Koshino, H.; Satoh, H. J. Chem. Inf. Model. 2007, 47, 1734–1746.

(42) Koichi, S.; Arisaka, M.; Koshino, H.; Aoki, A.; Iwata, S.; Uno, T.; Satoh, H. J. Chem. Inf. Model. **2014**, *54*, 1027–1035.

(43) Takahashi, S.; Yasuda, M.; Nakamura, T.; Hatano, K.; Matsuoka, K.; Koshino, H. J. Org. Chem. **2014**, *79*, 9373–9380.

(44) Mukhina, O. A.; Koshino, H.; Crimmins, M. T.; Kutateladze, A. G. *Tetrahedron Lett.* **2015**, *56*, 4900–4903.

(45) Koshino, H.; Satoh, H.; Yamada, T.; Esumi, Y. *Tetrahedron Lett.* **2006**, 47, 4623–4626.

(46) Takahashi, S.; Satoh, H.; Hongo, Y.; Koshino, H. J. Org. Chem. 2007, 72, 4578–4581.

(47) See Supporting Information.

(48) The specific rotation values of natural aromin and montanacin D (32) are $[\alpha]_{\rm D}^{22}$ +10.3 (*c* 0.25, CHCl₃) (ref 5) and $[\alpha]_{\rm D}^{26}$ +11.5 (*c* 0.12, CHCl₃), respectively. 4,8-Di-*epi* montanacin D (ref 9a) was precluded as a candidate (ref 47) of natural aromin because of the specific rotation value $[[\alpha]_{\rm D}^{26}$ +25.6 (*c* 0.12, CHCl₃)].

(49) The structure of aromin-A (ref 4) should also be reinvestigated because the compound was shown to be the C-20 epimer of aromin. Its absolute configuration at the left-half segment corresponding to C-15-C-32 is not reported.